

Plant–Insect Interactions in a Changing World

S. Pincebourde^{*1}, J. van Baaren[§], S. Rasmann[¶], P. Rasmont^{||},
G. Rodet[#], B. Martinet^{||} and P.-A. Calatayud^{** §§}

^{*}Institut de Recherche sur la Biologie de l'Insecte, UMR 7261, CNRS – Université François-Rabelais de Tours, Tours, France

[§]UMR-CNRS 6553 Ecobio, Université de Rennes 1, Université Bretagne-Loire, Rennes, France

[¶]University of Neuchâtel, Neuchâtel, Switzerland

^{||}University of Mons, Mons, Belgium

[#]AE, INRA, Avignon, France

^{**}UMR EGCE (Evolution, Génome, Comportement, Ecologie), CNRS-IRD-Univ. Paris-Sud, IDEEV, Université Paris-Saclay, Gif-sur-Yvette Cedex, France

^{§§}IRD c/o ICIPE, Nairobi, Kenya

^{*}Corresponding author: E-mail: sylvain.pincebourde@univ-tours.fr

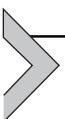
Contents

1. Introduction	2
2. Direct Effects of Climate Change on Plant–Insect Interactions	6
2.1 The Plant Side: Impact of Climate Change on Plant Defence Mechanisms	6
2.1.1 Effect of Elevated CO ₂ on Plant Defence and Resistance to Herbivore	7
2.1.2 Effect of Elevated Temperature on Plant Defence and Resistance to Herbivore	8
2.2 The Insect Side: Impacts of Climate Change on Herbivore Performance	10
2.2.1 Effect of CO ₂ Addition on Insect Performance	10
2.2.2 Effect of Temperature on Insect Performance	11
2.3 Impact of Climate Change on Pollinators	13
3. Indirect Effects of Climate Change on Plant–Insect Interactions	17
3.1 Plants Responses Matter for Insects	17
3.1.1 Impacts of Climate Change on the Plant Nutritional Quality for Herbivores	17
3.1.2 Cascading Effects of Changing the Plant Eco-Physiology on the Insect Microclimate	18
3.2 Biotic Interactions Matter for Insects	19
3.2.1 Thermal Traits Diverge Across the Different Components of a Multitrophic System	19
3.2.2 Synchronization Between Trophic Levels	22
3.2.3 Impacts at the Guild or at the Community Levels	24
4. Impact of Human Activities on Plant–Insect Interactions	24
4.1 The Main Anthropogenic Drivers Affecting Plant–Insect Interactions	24
4.1.1 Effects of Human Eco-Engineering	25
4.1.2 Evolutionary Responses to Accidental Anthropogenic Drivers	27
4.2 Impact of Pollution on Plant–Insect Interactions	28
4.2.1 Bottom–Up Effects of Pollution	29
4.2.2 Top–Down and Guild Effects of Pollution	30

5. Conclusion and Perspectives	30
Acknowledgements	32
References	32

Abstract

Global change is resetting the spatial and ecological equilibrium of complex co-evolutionary relationships between plants and their insect herbivores. We review the mechanisms at play in the responses of plant–insect interactions to global changes, including increased temperature and atmospheric CO₂ concentrations, modification of land use and pollution. We distinguish between the direct effects of global changes on each partner from the indirect impacts on insects via the responses of plants. The indirect effects include a change in the nutritional quality of the plant tissues for herbivore insects, as well as a change in the microclimatic conditions at the leaf surface. Pollinators are involved in a close symbiotic relationship with their favourite plants, and any depression caused by climate stress can lead to pollination deficit. Pollinators are, indeed, quite sensitive to global changes. Furthermore, although species are connected by trophic links, all species respond differently to global changes. We highlight that more research is needed to elucidate the plant-mediated indirect effects of climate change on insects. Then, other human activities, such as land transformations and release of pollutants, are likely to modulate these links between climate and plant–insect relationships. We argue that predicting the net effect of global change on plant–insect relationships requires a comprehensive understanding of the mechanisms that modulate the interaction strength between the plants and the insects, rather than on focusing on each partner individually.



1. INTRODUCTION

Earth is experiencing rapid changes in both climatic conditions and landscape structure. CO₂ concentration in the atmosphere has risen from about 280 ppm during pre-industrial times to the current 397 ppm. The atmospheric CO₂ concentration is predicted to reach up to 600 ppm towards the end of the 21st century if the rate of fossil fuel combustion does not change (IPCC, 2014). As a consequence, global average temperature would increase from 1.7°C to 6°C by the year 2100 (IPCC, 2014). In addition, an increase in the frequency of extreme events is expected (Hance, van Baaren, Vernon, & Boivin, 2007). Meanwhile, European landscapes have undergone a simplification through agricultural intensification leading to the removal of hedges, groves, woods and natural grasslands, and tending to reduce the effectiveness of ecosystem services such as biological pests control (Thies et al., 2011). Thus, even if their combined influences on the

ecosystems are still poorly valued, there is increasing evidence that working on both landscape properties and climate aspects can greatly advance our understanding of their impacts on plant–insect interactions (Tougeron, van Baaren, Burel, & Alford, 2016). These changes in climate and landscapes are disrupting the energetic budget of plants, ultimately affecting their eco-physiological responses against insect herbivores. Therefore, global change is resetting the spatial and ecological equilibrium of complex co-evolutionary relationships between plants and their insect herbivores (Coley, 1998).

Humans disturb the ecosystems in which species live and evolve, leading to ecological and evolutionary consequences for plant–insect interactions (Mitter, Farrell, & Futuyma, 1991; Rundle & Nosal, 2005). Intentionally, humans create totally new environments, such as agriculture fields and cities, in which fauna and flora are subjected to new selection pressures. These new environments are maintained only through human actions. On the other hand, accidentally, humans disturb natural ecosystems due to proximity, needs, pollution and unexpected consequences of their actions. These effects can occur over large areas, at great distances from their anthropogenic origin. Humans are ingenious but they seem to be impotent when faced with the fact that all their innovations are accompanied with the emergence of new accidents (Virilio, 2005). Global climate change is probably the greatest accident. Overall, humans influence ecological processes in three ways: (1) by altering environmental conditions through modifications and transformations of the landscape, (2) by creating new niches in anthropized environments and (3) by rearranging ecological communities through species introduction or removal.

Since more than 500 million years ago, plants and insect herbivores have been engaged in an evolutionary arms race that has been the source of major diversification events (e.g., Currano, Labandeira, & Wilf, 2010; for a review see Chapters: Plant–Insect Interactions: a Paleontological and an Evolutionary Perspective and Evolution of Plant–Insect Interactions: Insights From Macroevolutionary Approaches in Plants and Herbivorous Insects by Kergoat, Meseguer, Jousselin, 2016), which has resulted in plants covering more than half of the biodiversity described today (Strong, Lawton, & Southwood, 1984). Due to their sessile nature, plants evolved sophisticated strategies to protect themselves from herbivore attacks, including structural and chemical means (Schoonhoven, van Loon, & Dicke, 2005). In turn, insects responded to this incredible chemical diversity by evolving detoxification mechanisms (Feyereisen, 1999; Heckel, 2014; for a review see chapter: From Plant Exploitation to Mutualism by Lieutier et al.,

2016), behavioural avoidance mechanisms (Dussourd & Denno, 1991) and temporal and spatial specialization for different plant organs (Jaenike, 1990). Because the equilibrium of the complex interactions between plants and insects depends on environmental conditions, current global change is likely to induce profound changes in the strength of the plant–insect interactions (Tylianakis, Didham, Bascompte, & Wardle, 2008).

Species are linked by trophic links (see Chapters: Food Webs and Multiple Biotic Interactions and The Plant as a Habitat for Entomophagous Insects by Kaiser et al., 2016). In a given community, plants are attacked by herbivores which are themselves attacked by predators and parasitoids. Insect herbivores such as aphids contain different obligatory and facultative endosymbiotic bacteria (see chapter: Influence of Microbial Symbionts on Plant–Insect Interactions by Giron et al., 2016). In ecosystems, these multitrophic interactions are the result of a long co-evolutionary process that unfolds within a particular environment. Any modification occurring at one trophic level will have repercussions on the others. Indeed, each species of the multitrophic network has its own tolerance to temperature, CO₂ concentration or humidity. The preferendum may differ between species and trophic levels, causing various responses to climate change. For example, *Trichogramma chilonis* Ishii 1941 (Hymenoptera: Trichogrammatidae) has an optimal response at temperatures between 25°C and 35°C, whereas it is between 30°C and 35°C for its competitor *Cotesia vestalis* (Haldy, 1834) (Hymenoptera: Braconidae) (Reddy et al., 2002). During the quaternary period, global warming induced distribution shifts with different intensities and speed according to the species: the individual responses of each population or species produced new communities (Graham & Grimm, 1990). To predict the impact of climate change at the community level, it is therefore necessary to understand what will happen to every single species, at each trophic level (guild level), at the level of the synchronism between trophic levels and finally at the community level (Tylianakis et al., 2008).

A well-discussed case of how climate change might impact plant–insect interactions is the effect of environmental temperature. Global warming causes variation in the composition of the plants and their herbivore communities, particularly through differential colonization abilities from more sessile plants to more mobile animal herbivores (Burrows et al., 2011; Parmesan, 1996; Pateman, Hill, Roy, Fox, & Thomas, 2012; Pauli et al., 2012; Stange & Ayres, 2001), and also through shifts in phenological events such as the flowering date or emergence date of overwintering insects (Parmesan & Yohe, 2003). Such phenological mismatch should have

profound effects on the co-evolved plant–herbivore interactions (Harrington, Woiwod, & Sparks, 1999). These effects will be largely mediated by plant defence traits (Rasmann & Pellissier, 2015). Nevertheless, the magnitude and direction of change still remains to be fully elucidated, since the mechanisms at play are highly context dependent and variable (Zavala, Nabity, & DeLucia, 2013; Zvereva & Kozlov, 2006).

Here, we review the mechanisms by which global climate change and human activities impact the way plants and insects interact with each other. We do not pretend to exhaustively cover the various effects of global change on plant–insect interactions (for a review see Björkman & Niemelä, 2015).

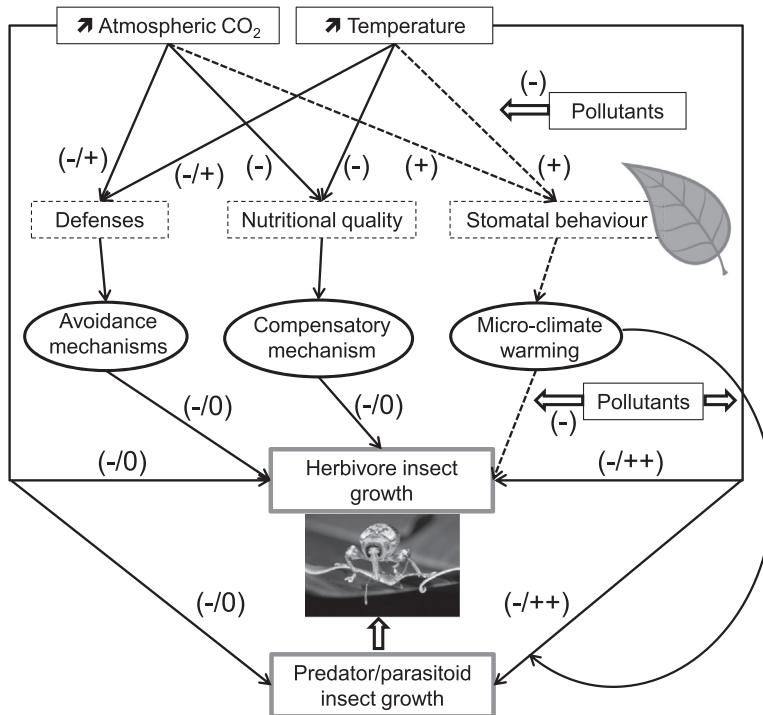
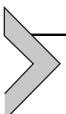


Figure 1 Diagram illustrating the complexity of the multiple interactions between some of the abiotic variables associated with global change (CO₂, temperature, pollution), some plant key processes (structural and chemical defences against herbivores, nutritional quality of plant tissues, stomatal ecophysiology), some herbivore insect responses (avoidance behaviour, compensatory responses) and the third trophic level, the predators and parasitoids. Signs (-/+0) indicate the possible effect, negative, positive or no effect, respectively. This diagram is not exhaustive, and other mechanisms may play a significant role.

Our aim is to emphasize the mechanisms behind the responses of plants and insects, and how they interact (Fig. 1). We specifically focus on the role of rising atmospheric CO₂ concentration and increasing temperature. The direct effects on both the plants and the insects are reviewed separately from the indirect effects of these factors on the insects via their influence on the plants. In particular, we highlight that more research is needed to elucidate the plant-mediated indirect effects of climate change on insects. Finally, we detail the effects of human activities on plant–insect relationships by focusing on land use and on pollution. We argue that predicting the net effect of global change on plant–insect relationships requires a comprehensive understanding of the mechanisms that modulate the interaction strength between the plants and the insects, rather than focusing on each partner individually.



2. DIRECT EFFECTS OF CLIMATE CHANGE ON PLANT–INSECT INTERACTIONS

2.1 The Plant Side: Impact of Climate Change on Plant Defence Mechanisms

The arms race between plants and insects has led to the evolution of plant mechanisms to minimize damage by insects (Futuyma & Agrawal, 2009; Mitter et al., 1991), including the deployment of chemical and physical defence traits (Schoonhoven et al., 2005). More specifically, plant resistance against herbivores is mediated by structural attributes, such as thick and tough epidermal layers, spines, trichomes, as well as toxic phytochemicals (also called secondary metabolites) that can inhibit, intoxicate and/or kill the herbivore (Schoonhoven et al., 2005). The main classes of secondary metabolites include the terpenoids, phenolics and alkaloids, and also other nitrogen- or sulphur-containing compounds such as the glucosinolates in the Brassicaceae or sugar-bound molecules such as the iridoids in the Plantaginaceae (Rosenthal & Berenbaum, 1991).

In addition to directly deploying physical and chemical defences, it has become a general agreement that plants can benefit from attracting predators near the site of herbivore attack (Dicke & Baldwin, 2010). More specifically, indirect defences involve the production of features that provide shelter (e.g., domatia), reward (e.g., extrafloral nectar) or information on herbivore presence, such as the release of volatile organic compounds (VOCs), for the natural enemies of herbivores (Kessler & Heil, 2011). These defence traits mediate herbivore–enemy interactions by increasing herbivore suppression,

and ultimately potentially increasing plant fitness (Romero & Koricheva, 2011; Schmitz, Hamback, & Beckerman, 2000).

While most of the variations in plant's defence arsenal are attributed to a variety of genetic, ontogenetic and phenologic components, including evolutionary history (Futuyma & Agrawal, 2009), plants also have to constantly cope with variations in environmental conditions, generally imposing a reshuffling of the carbon/energy balance, and thus the allocation between growth and defence (Coley, Bryant, & Chapin, 1985; Herms & Mattson, 1992). In the next section, we will discuss the general patterns emerging from several years of research on the two major climate change drivers – elevated CO₂ and temperature – on plant direct and indirect defences and how they might affect plant's resistance to herbivore attack.

2.1.1 Effect of Elevated CO₂ on Plant Defence and Resistance to Herbivore

The effect of elevated CO₂ on insect herbivores is mainly mediated by changes in plant chemistry (Cornelissen, 2011; Lincoln, Fajer, & Johnson, 1993). An overall conclusion drawn from summarizing the effect of CO₂ addition on plants suggests that changes in primary metabolism are in fact relatively predictable, whereas changes in plant's secondary metabolite production, and their corresponding effects on plant direct and indirect defences, are highly variable (Bidart-Bouzat & Imeh-Nathaniel, 2008), with subsequent strong variation in herbivore performance (Coviella & Trumble, 1999; Robinson, Ryan, & Newman, 2012). Elevated CO₂ typically increases the carbon:nitrogen ratio (Robinson et al., 2012). In addition, it has dramatic but variable impact on the general leaf secondary chemistry (Bidart-Bouzat & Imeh-Nathaniel, 2008; Cornelissen, 2011; Lindroth, 2010; Stiling & Cornelissen, 2007). Perhaps the only exception is the production of VOCs, which was consistently shown to increase in concentration in the headspace surrounding the plant under elevated CO₂ conditions (Peñuelas & Staudt, 2010). Nevertheless, such increase does not automatically translate into higher herbivore suppression by predators in the field, and again, context dependency applies when analyzing the community-wide impacts of elevated CO₂ (Facey, Ellsworth, Staley, Wright, & Johnson, 2014). This variation poses a clear challenge for developing credible predictions of how further CO₂ addition into the atmosphere will impact the plant resistance against herbivores. Therefore, one has to move away from the highly variable secondary metabolism and

look at other, more predictable traits. For instance, a relatively small literature which is rapidly gaining attention shows a direct effect of elevated CO₂ on plant hormones (phytohormones), such as jasmonic acid (JA) and salicylic acid (SA) (see below).

Plants attacked by herbivores generate specific hormonal cascades which ultimately elicit downstream changes in plant biochemistry and secondary metabolism through complex signalling networks (Browse & Howe, 2008; Wu & Baldwin, 2010). Both JA and SA are involved in the orchestration of plant defence after herbivore attack (Farmer, Almérás, & Krishnamurthy, 2003; Thaler, Agrawal, & Halitschke, 2010). Exposure to elevated CO₂ can modulate hormone production, which might explain some of the variations in the observed responses of allelochemicals (Ode, Johnson, & Moore, 2014; Zavala et al., 2013). To summarize, the emerging picture suggests that (1) elevated CO₂ down-regulates the constitutive herbivore-induced expression of several key genes associated with the JA pathway, resulting in an increased herbivore damage in field and laboratory conditions (Sun et al., 2011; Zavala, Casteel, DeLucia, & Berenbaum, 2008); (2) because of the inherent cross-talks between JA- and SA-signalling pathways (Thaler, Humphrey, & Whiteman, 2012), elevated CO₂ enhances induced defences derived from the SA-signalling pathway (Ghasemzadeh, Jaafar, & Rahmat, 2010), such as the pathogenesis-related protein (Sun et al., 2011) and (3) the differential responses among phytohormonal pathways provide new insights into how elevated CO₂ modulates plant defences against herbivory.

2.1.2 Effect of Elevated Temperature on Plant Defence and Resistance to Herbivore

Temperature affects the plant physiological processes, growth, reproduction and survival (Jamieson, Trowbridge, Raffa, & Lindroth, 2012; Julkunen-Tiitto, Nybakken, Randriamanana, & Virjamo, 2015). In addition, warming modifies the biosynthesis of plant secondary metabolites (Bidart-Bouzat & Imeh-Nathaniel, 2008; Loreto & Schnitzler, 2010). A general trend was reported for the responses of phytochemicals to enhanced temperature (Zvereva & Kozlov, 2006). Phenolics tended to decrease, whereas terpenes tended to increase at higher temperatures. However, any generalization remains difficult since several examples showed no, or even the opposite effects of elevated temperature on phenolic-based compounds, alkaloids, terpenoids, or glucosinolates (Bidart-Bouzat & Imeh-Nathaniel, 2008; Julkunen-Tiitto et al., 2015). When looking at the effect of increased

temperature on the production of VOCs, the picture seems to be clearer. VOC production generally increases with temperature up to the physiological maximum of the plant, beyond which enzymatic degradation inhibits emission (Guenther, Zimmerman, Harley, Monson, & Fall, 1993). The mechanisms of increased emission reside particularly on enhanced enzymatic activities, and increased VOC vapour pressure (Peñuelas & Llusià, 2003). Interestingly, Peñuelas and Llusià, (2003) suggested that increased VOC production should not only be related to enhanced biotic interactions (pollinator and/or predator attraction), which might be also less predictable (de Sassi, Staniczenko, & Tylianakis, 2012; Tylianakis et al., 2008), but also to an increased thermal tolerance for plants. In other words, increased temperatures favour VOC emissions, which in turn enable the plant to better withstand the heat stress.

Another approach for studying the effect of temperature changes on plant defences and plant–insect interactions is to use elevation gradients as natural source of variation in biotic and abiotic factors (Körner, 2007; Rasmann, Alvarez, & Pellissier, 2014; Rasmann, Pellissier, Defossez, Jactel, & Kunstler, 2014). Recent examples show that high-altitude adapted *Plantago lanceolata* L. or *Vicia sepium* L. genotypes produce lower levels of secondary metabolites (iridoid glycosides or VOCs, respectively). While such decline could be attributed to a reduction in herbivore pressure at high elevation, colder temperatures also inhibit iridoid glycoside production in *P. lanceolata* (Pellissier, Roger, Bilat, & Rasmann, 2014). A general decline in plant resistance at high altitude seems to be the rule (Pellissier et al., 2012; for exceptions see Rasmann, Alvarez, et al., 2014; Rasmann, Pellissier, et al., 2014). Specifically, we can hypothesize a general decline of plant defences at high elevation for the predominant plants in the community, while the rarest plants benefit from having their slow growing leaves well protected (Fine, Mesones, & Coley, 2004; Pellissier et al., 2016).

According to predictions, climate warming will reshuffle plant and herbivore communities along altitudinal gradients through high elevation colonization mismatch (Pauli et al., 2012). Therefore, we suggest that high elevation plants will be more exposed to herbivore pressure in the upcoming years (Rasmann, Alvarez, et al., 2014; Rasmann, Pellissier, et al., 2014). Plants' tolerance to such predicted increase in herbivory will be the outcome of several factors, including genetic (and epigenetic) variations, the ability to change their phenotype rapidly in the presence of herbivore attack (i.e. phenotypic plasticity), and ultimately the rate to which plants can adapt (Rasmann & Pellissier, 2015). In other words,

climate warming will indirectly impose a strong habitat-specific selection pressure on plants, in turn favouring evolution for novel defence syndromes at high elevation.

2.2 The Insect Side: Impacts of Climate Change on Herbivore Performance

2.2.1 Effect of CO₂ Addition on Insect Performance

The direct effect of changing CO₂ concentration in the atmosphere on the eco-physiology of insects has been largely overlooked till date. Recently, Kerr, Phelan, and Woods (2013) showed that the developmental rate of *Manduca sexta* Linnaeus 1763 caterpillars was only slightly affected when reared under high (above 1200 ppm) CO₂ concentrations. More importantly, rising atmospheric CO₂ concentration may be linked to changes in the acidification of insect body fluids. This may be especially true for the eggs that cannot ventilate like the larvae and adult stages, but can only exchange gases via passive diffusion (Woods, Bonnecaze, & Zrubek, 2005). Nevertheless, exposing *M. sexta* eggs to high CO₂ concentrations for several hours did not influence the pH in the egg yolk (Kerr et al., 2013). Undoubtedly, more experimental evidence is needed to generalize these findings.

Rising atmospheric CO₂ concentration may be more likely detrimental to insect species that already live in hypercarbic environments such as cambium miners, stem borers and insects that induce large galls (Pincebourde & Casas, 2016). These endophagous insects live deep into the plant tissues and gases diffuse slowly between outside the plant and the insect location. Therefore, an increase in atmospheric CO₂ concentration may slow down the diffusion of CO₂ outward, enhancing the risk of reaching dangerous asphyxiating levels (Pincebourde & Casas, 2016). Indeed, the hypercarbic conditions in the galleries of these insects coincide with hypoxia, i.e. low oxygen levels. But, again, a lack of knowledge on the direct physiological effects of CO₂ on insect herbivores precludes any firm conclusion.

Finally, changes in CO₂ could also affect the behaviour of insects. Particularly, several soil-dwelling herbivores utilize root-emitted CO₂ as a cue for finding their hosts (Nicolas & Sillans, 1989). Several species can be quite sensitive to variations in CO₂ levels, as they can detect CO₂ concentration increase as small as 0.003% and locate their host plant from a distance of 20 cm (Doane, Lee, Klingler, & Westcott, 1975). This peculiarity makes these insects especially sensitive to rising atmospheric CO₂ concentration.

2.2.2 Effect of Temperature on Insect Performance

Most insects are ectotherms and temperature variations were shown to directly influence their eco-physiology including: metabolic rate, growth, feeding rates and life-history traits, such as fecundity and longevity (Angilletta, 2009; Bale et al., 2002). Generally, the thermal response of insects follows the classic thermal performance curve. An insect can increase the performance above a threshold (minimal critical temperature, CT_{\min}) up to the optimal temperature, after which performance decreases sharply and reaches the upper threshold for performance (maximal critical temperature, CT_{\max}) (Angilletta, 2009). Therefore, a warming climate should generally improve the eco-physiological performance of insects, as long as the temperature does not surpass the temperature optimum, and does not reach the CT_{\max} . The ecological consequences of temperature-mediated improved performance include the expansion of species distribution ranges, as new thermal niches appear northwards (Parmesan, 2006; Parmesan & Yohe, 2003), and the increase of insect population outbreaks (Jepsen, Hagen, Ims, & Yoccoz, 2008). Understanding these processes, however,

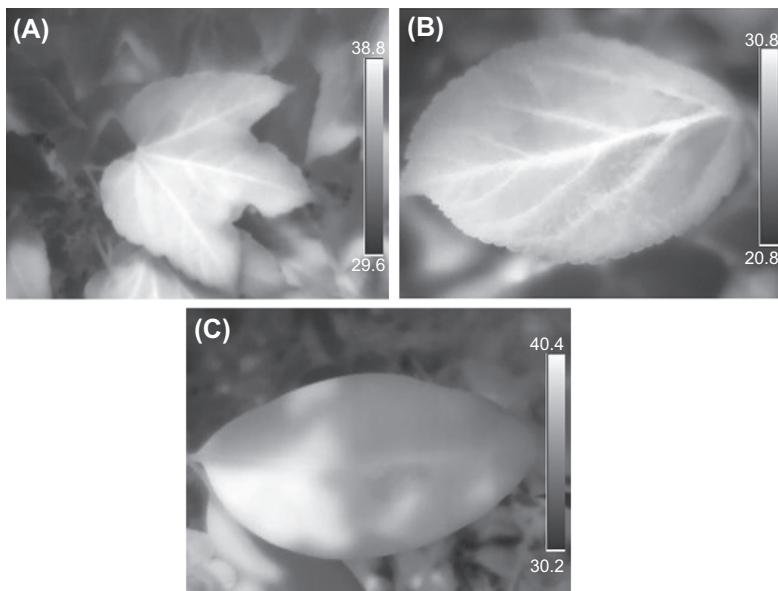


Figure 2 Thermographic images of leaf surfaces exposed to solar radiation for (A) grape ivy (France), (B) apple (France) and (C) *Clusia* sp. (French Guiana). During these measurements with an infrared camera, air temperature was 28.6°C, 24°C and 29.2°C, respectively.

necessitates clarifying the predictions of when and where the temperature falls above the optimal temperature and reaches the CT_{max} of species (Sunday et al., 2014). Specifically, this requires the determination of the exact temperature experienced by insects at the leaf surface or within the leaf tissues (Pincebourde & Woods, 2012).

The temperature at the surface of plants can deviate from ambient air temperature (Fig. 2). For instance, the leaf temperature excess (i.e. temperature deviation between leaf surface and ambient air) can be up to 10°C in temperate plants (Cook, Dixon, & Leopold, 1964; Pincebourde, Sinoquet, Combes, & Casas, 2007) and even up to 20°C in alpine plants in full sunlight (Linacre, 1967). By contrast, in the arid ecosystem of Arizona (USA), *Datura* leaves are colder than the surroundings during the day (Potter, Davidowitz, & Woods, 2009). The developmental and tolerance thresholds of *Manduca sexta* eggs are adapted to the *Datura* leaf temperature patterns, as they would die if they experience the high desert ambient air temperature (Potter et al., 2009). Generally, the excess temperature of leaves decreases when ambient air increases (Linacre, 1967; Michaletz et al., 2015; Pincebourde & Woods, 2012). The shape of the relationships between leaf temperature and ambient air temperature, however, may depend on the plant system and on the spatial scale at which the process is investigated (Pincebourde, Murdock, Vickers, & Sears, 2016).

The degree to which leaf temperature influences insects depends on their body size. The main reason is purely physical: any organism small enough to remain totally embedded into the leaf boundary layer is subjected to leaf temperature directly (Kaspari, Clay, Lucas, Yanoviak, & Kay, 2015). By contrast, organisms larger than the leaf boundary layer mainly depend on the ambient conditions (Woods, 2013). Across their ontogeny, most herbivore insects experience leaf temperature fluctuation during the first stages of their development until they reach a body size, beyond which their thermal budget shifts as they are influenced by ambient air conditions. For example, early instars of the *Manduca sexta* caterpillar experience temperature about 5°C below ambient air at the *Datura* leaf surface, while the last larval stage reaches body temperatures that are several degrees above ambient air (Woods, 2013).

Understanding how warmer climates affect insects thus requires quantifying the effect of global warming on leaf surface temperatures. From an evolutionary perspective, the convergence of mean leaf temperature at all latitudes towards a value of $\sim 22^\circ\text{C}$ (Helliker & Richter, 2008) indicates that climate change may have a relatively small effect on leaf surface

temperature, as long as the whole plant can compensate. Nevertheless, it is not clear how global change will cause changes at the leaf surface, and several hypotheses were developed recently (Pincebourde & Woods, 2012). Depending on the level of adaptation to environmental fluctuations that are larger in the temperate zone compared to lower latitudes, the amplitude of warming may be buffered or by contrast be reported to the leaf surface temperature—more empirical research is needed. In addition, the ability of arthropods for behavioural thermoregulation could modify the leaf surface temperature pattern they experience (Pincebourde & Suppo, 2016). Spider mites, for example, make use of the within-leaf surface thermal heterogeneity to thermoregulate and keep track of the optimal temperature for their development (Caillon, Suppo, Casas, Woods, & Pincebourde, 2014). However, warming homogenizes leaf surface temperatures, thereby disrupting the behavioural thermo-compensation of mites, and ultimately leading them to overheat (Caillon et al., 2014).

2.3 Impact of Climate Change on Pollinators

Recently, climate change was pointed out as one of the major causes of regression of insect pollinator populations. While other groups may be involved, most of the pollination service is provided by the ~20,000 bee species (Michener, 2007; Proctor, Yeo, & Lack, 1996). Interestingly, these insects, and in particular bumblebees, generally show an elaborated endothermic behaviour, which allowed them to colonize a vast number of habitats (Heinrich, 1979; Owen, Bale, & Hayward, 2013). This specialized endothermy, however, exposes bumblebees to climate risks.

The modelling of eco-climatic envelope for bumblebee species indicated that many species will be significantly rarefied due to global warming (Rasmont et al., 2015). The climatic risk is more or less accentuated depending on the scenarios considered by IPCC (2007) (Fig. 3). In Europe, the distribution of 30, 47 and 53 species among the 69 bumblebee species will be reduced according to the scenario SEDG, BAMBU and GRAS, respectively (Settele et al., 2005). The population disruptions due to climate change can be severe. Specifically, for areas such as Madrid, Paris, London, and Helsinki, only one to three species can survive while several dozen species were present in the 20th century (Rasmont et al., 2015). By contrast, pollinators may find refuges in mountain areas which will conserve their current diversity. Therefore, population movements will likely determine the survival of these species. Comparable trends are expected for North America, except that the Northern margin of bumblebee distribution will

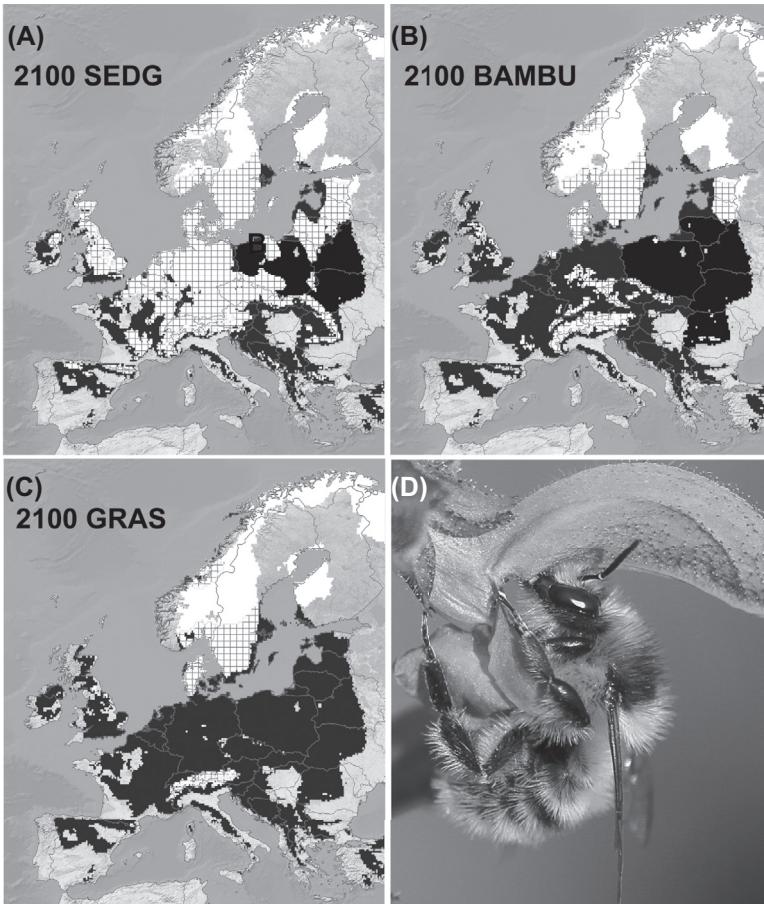


Figure 3 Projected suitable climatic area of *Bombus sylvarum* in 2100 in Europe. (A) SEDG scenario, (B) BAMBU scenario, (C) GRAS scenario and (D) *Bombus sylvarum* visiting a flower. Photo by J. Carteron. Areas in black will be lost; hatched areas will be conserved; areas in white will be gained. After Rasmont, P., Franzen, M., Lecocq, T., Harpke, A., Roberts, S.P.M., Biesmeijer, K., ..., Schweiger O. (2015). Climatic risk and distribution atlas of European bumblebees. *Biorisk*, 10, 1–246. <http://dx.doi.org/10.3897/biorisk.10.4749>.

not shift as much as the Southern limits (Kerr et al., 2015). North American taxa may have smaller dispersion ability than European species. Indeed, the postglacial history of species in Europe may have induced a strong selection of the most mobile taxa (Hewitt, 1999; Lecocq et al., 2013).

Some isolated mesoclimatic zones in southern European regions can play the role of ‘Noah’s Ark’ by becoming a source-bumblebee population for

this entire geographic region (Rasmont et al., 2015). Unfortunately, this perspective could be compromised by the genetic drift of the small surviving isolates. Indeed, the mechanism of sex determination in bumblebees may contribute to their decline in a warmer world. The male, normally haploid, is determined by the homozygosity of a small number of loci (Duchateau, Hoshiba, & Velthuis, 1994). During inbreeding, a large number of diploid males emerge with a drastically reduced fertility. Therefore, the monomorphism of these loci induced by genetic drift could depress the reproductive potential (Whitehorn, Tinsley, Brown, Darvill, & Goulson, 2009). Only large mountain areas, which will shelter large and genetically diverse residual populations, could ensure the survival of these pollinator species in the future.

The modelling studies of Kerr et al. (2015) and Rasmont et al. (2015) are mainly based on temperature averages. These models do not include extreme climatic events such as heat waves which will increase in frequency and duration in the next decades (Meehl & Tebaldi, 2004; Robinson, 2001) and which can cause local extinctions of some bumblebee species (Rasmont & Iserbyt, 2012). Martinet, Lecocq, Smet, and Rasmont (2015) showed that a typical heat wave temperature (40°C in temperate regions) rapidly kills the tested specimens (Fig. 4). Some bumblebee species, however, are more sensitive to thermal stress than others (e.g., arctic and top mountain species). Moreover, critical thermal limits vary according to altitudinal distribution of bumblebees (Oyen, Susma, & Dillon, 2016). By contrast, a species such as *Bombus terrestris* (Linnaeus, 1758) seems to tolerate well heat wave conditions. The particular thermal tolerance of this species may explain its recent Northward expansion in Europe despite the repeated heat waves (Martinet et al., 2016).

Some wild bee species are linked to hot and dry climates (Michener, 1979). For instance, the honeybee *Apis mellifera* Linnaeus 1758 or *Xylocopa varipuncta* Patton 1879 are able to fly in very hot conditions (Heinrich, 1979; Heinrich & Buchmann, 1986). Numerous wild bees are living in desert and other dry habitats, with their maximum diversity in these environments (Patiny & Michez, 2007). As of today, however, we have no evaluation of the thermal tolerance in these thermophilic taxa or on their resistance to climate change (Nieto et al., 2014, p. 84). For a small number of species, global warming is the best explanation for their northward expansion, e.g., *Xylocopa (Koptortosoma) pubescens* Spinola 1838 which is now present in Europe (Terzo & Rasmont, 2014).

Beyond the fate of pollinator insects during climate change, the question of pollination service is crucial. Both the plant and its pollinators may respond differently to climate change. For example, the large shift towards

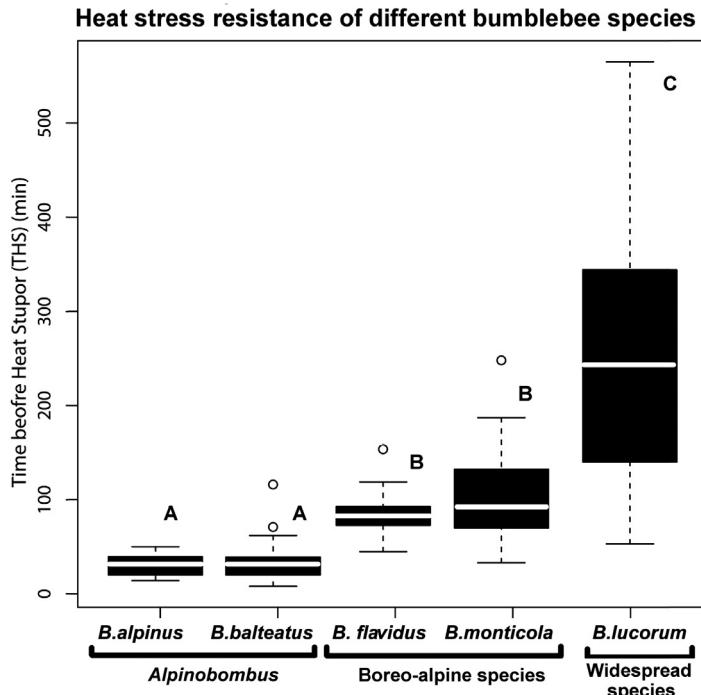


Figure 4 Boxplots of the time before heat stupor for six bumblebees species (*Bombus* sp.). (A) Arcto-alpine species: *Bombus alpinus* and *Bombus balteatus*, (B) boreo-alpine species: *Bombus flavidus* and *Bombus monticola* and (C) widespread species: *Bombus lucorum*. Circles are extreme values. After Martinet, B., Lecocq, T., Smet, J., & Rasmont, P. (2015). A protocol to assess insect resistance to heat waves, applied to bumblebees (*Bombus Latreille, 1802*). PLoS One, 10, e0118591.

the North of orchards relative to the small shift of the associated pollinators may generate an extensive pollination deficit area (Polce et al., 2014). In general, pollinators are involved in a close symbiotic relationship with their favourite plants, and any depression caused by climate stress can lead to pollination deficit (Franzén & Ockinger, 2012). This deficit can significantly reduce the reproductive potential of both domesticated and wild plants. In turn, this reduction can lower biodiversity by generating food deficiency for the species associated with these plants. This interaction chain still escapes our modelling capabilities.



3. INDIRECT EFFECTS OF CLIMATE CHANGE ON PLANT–INSECT INTERACTIONS

3.1 Plants Responses Matter for Insects

3.1.1 *Impacts of Climate Change on the Plant Nutritional Quality for Herbivores*

Rising atmospheric CO₂ concentration influences the development of herbivores indirectly by inducing changes in the plant nutritional quality (Lincoln et al., 1993). In general, plants growing in a CO₂-enriched atmosphere have higher levels of non-structural sugars, which are beneficial to insect herbivores (Bezemer & Jones, 1998). Nevertheless, higher CO₂ levels also lead to a lower concentration of proteins (Ehleringer, Cerling, & Dearing, 2002; Whittaker, 2001) and some amino acids (Docherty, Wade, Hurst, Whittaker, & Lea, 1997) in leaves. In addition, the concentration of secondary compounds, which are toxic to herbivore insects, increases in a CO₂-enriched atmosphere (Bidart-Bouzat, Mithen, & Berenbaum, 2005; Cornelissen, Stiling, & Drake, 2003; Stiling & Cornelissen, 2007). Therefore, the nutritional quality of plants is lower when grown under high CO₂ levels compared to current concentrations. Negative impacts of increasing CO₂ levels were also found on the development rate and survival of phytophagous insects (Smith & Jones, 1998; Whittaker, 2001). Herbivore insects may develop compensatory mechanisms by increasing their food intake quantitatively or by enhancing assimilation efficiency (Barbehenn, Karowe, & Chen, 2004; Stiling & Cornelissen, 2007). For example, leaf miner caterpillars compensate for the lower quality of their plant food by increasing the amount of leaf tissues eaten (Cornelissen et al., 2003; Salt, Brooks, & Whittaker, 1995). Compensatory mechanisms were not found in all species (Smith & Jones, 1998), however, and even when compensation occurs it does not cover all the negative impacts of low quality food (Hunter, 2001).

Temperature changes can also influence the nutritional quality of host plants. The net effect is highly plant species specific. A long-term increase in temperature induces a decrease in the water content of leaves (Williams, Norby, & Lincoln, 2000). In herbaceous, warming induces an increase in nitrogen concentration of plant tissues (Bezemer & Jones, 1998). By contrast, in woody species, warming negatively impacts the nitrogen concentration and increases the concentration of tannins (Dury, Good, Perrins, Buse, & Kaye, 1998). The concentration of sugar can also be affected by the increase in temperature (Morison & Lawlor, 1999).

Interactive effects of temperature increase and CO₂ concentration change were reported, but the net effect varies according to the insect–plant system (Zvereva & Kozlov, 2006). In some systems, only one of the two factors has an impact on the development rate of the insect, either negative (Williams et al., 2000) or positive (Buse & Good, 1996). In other systems, however, temperature and CO₂ interact such that the negative effect of one is suppressed by the positive effect of the second (Fajer, Bowers, & Bazzaz, 1991). Finally, in most cases, compensatory effects happen resulting in the compensation of the negative effect from one factor by the positive effect of the second (Johns, Beaumont, & Hughes, 2003; Johns & Hughes, 2002). Overall, the predicted negative effects of CO₂ elevation on herbivores are likely to be mitigated by temperature increase (Zvereva & Kozlov, 2006), but more experimental data are needed on different biological systems to generalize these findings (Robinson et al., 2012).

3.1.2 Cascading Effects of Changing the Plant Eco-Physiology on the Insect Microclimate

When feeding on their host plant, insects modify the plant tissues physically and/or physiologically. In turn, these modifications can induce shifts in the insect microclimate – an effect called the physical feedback of herbivory (Pincebourde & Casas, 2006a). This indirect effect is especially prominent in endophagous insects such as leaf miners and gallers. These organisms alter the structure and the properties of the leaf (Pincebourde & Casas, 2016). In general, the outcome of these modifications is an increase in the microclimatic temperature for the insect herbivore. For example, the leaf miner *Phyllonorycter blancardella* (Fabricius, 1781) (Lepidoptera: Gracillariidae) induces stomatal closure in apple leaf tissues (Pincebourde, Frak, Sinoquet, Régnard, & Casas, 2006) and generates white patches at the leaf surface (Pincebourde & Casas, 2006a). These two modifications contribute to an elevation of the temperature within the mine by up to 12°C above ambient air (Pincebourde & Casas, 2006b, 2015). These elevated temperatures can speed up the insect development, but this strategy is at risk during extreme climatic events such as heat waves (Pincebourde & Casas, 2015). During heat waves, the mosaic of favourable and risky microclimates is reshuffled (Pincebourde et al., 2007).

External feeders also induce variations in leaf surface temperatures during their feeding activity. These variations are in the order of few degrees, and are generally quite local around the leaf portion attacked by the herbivore (Nabity, Hillstrom, Lindroth, & DeLucia, 2012). These temperature

changes at the leaf surface are the consequence of alteration in stomatal conductance and evapotranspiration rates. Nevertheless, little is known on these interactive effects during warming. Indeed, insect herbivores tend to increase their feeding rate in a warmer environment (see above), suggesting that more local impacts on leaf transpiration rates are expected, thereby reinforcing the general warming effect. Overall, the plant responses to climate change should have consequences on the microclimatic conditions at the leaf surface. Many parameters impact the leaf heat budget, including its size and shape, stomatal conductance and its orientation towards the sun (Gates, 1980). Stomata have an important role in the plant responses: they regulate the overall energy and gas budget of the plant, and they react to several environmental (e.g., climatic variables) and endogenous factors (e.g., plant water status). Climate change can be expected to induce variations in these factors, with likely cascading effects on surface temperatures (Pincebourde & Woods, 2012). Finally, changing plant architecture may also partially buffer the amplitude of warming for insects at the leaf surface, but this effect seems to be quite limited given the magnitude of warming (Saudreau et al., 2013).

3.2 Biotic Interactions Matter for Insects

Species are linked by trophic links. Therefore, a series of indirect effects are expected to cascade through the entire food web, thereby increasing the difficulty to anticipate the effects, especially in complex multitrophic systems (Tylianakis et al., 2008).

3.2.1 Thermal Traits Diverge Across the Different Components of a Multitrophic System

3.2.1.1 Development Time and Growth Rate

The speed at which organisms respond to climate change is partially determined by their generation time. In general, temperature increases the development rate of plants and insects inside their tolerance range. For plants, the period of vegetation increases in length, allowing insects to feed on them for a longer period throughout the year. Some multivoltine species can realize an additional generation per year due to both the longest period of vegetation and their shorter development time. For example, in the European grapevine moth *Lobesia* (*Lobesia*) *botrana* (Denis & Schiffermüller, 1775) in South-West of France, Marchal and Feytaud (1911) observed three generations of this insect in 1910, but a century later Martin-Vertedor, Ferrero-Garcia, and Torres-Vila (2010) noted a fourth generation at the end of fall. The increased

reproduction rate influences the population dynamics at the other trophic levels. The parasitism rate of the eastern spruce budworm, *Choristoneura fumiferana* Clemens 1865 (Lepidoptera: Tortricidae) by its parasitoids decreases when the temperature increases (Harrington, Fleming, & Woiwod, 2001). By contrast, the predation rate of the seven-spot ladybird, *Coccinella septempunctata* Linnaeus 1758 on the aphid *Acyrtosiphum pisum* (Harris, 1776) increases at high temperatures. The impact of temperature increase cannot be generalized without the understanding of the specific thermal responses.

3.2.1.2 Metabolic Rate, Longevity and Fecundity

These traits are driven by temperature in natural enemies such as parasitoids, thereby modifying the parasitism rate and then the impact of phytophagous insects on plants. An increase in temperature increases metabolism and activity (May, 1979). For example, the walking speed of the parasitoid *Aphelinus asychis* Walker 1839 (Hymenoptera: Aphelinidae) increases with temperature (Mason & Hopper, 1997). These traits are associated with an increase in the instantaneous rate of parasitism. However, the increased metabolic rate also results in a reduced longevity due to the accelerated use of energy (Huey & Stevenson, 1979; Trotta et al., 2006). Therefore, the impact at the life span level is difficult to determine. In addition, above a given temperature threshold, the negative effects of high temperature exceed the advantages. This complexity of the relationship between traits at the individual scale is found at each trophic level. An increase of temperature can increase the fecundity of the phytophagous insect and decrease that of the parasitoid, or vice versa.

3.2.1.3 Sex Allocation

Temperature can influence sex allocation in insects. This was observed for arrhenotokous hymenopteran parasitoids, in which unfertilised eggs develop into haploid males and fertilised eggs develop into diploid females. Females are able to allocate male or female eggs in one host according to internal or external factors, thus affecting the sex ratio. For example, the sex ratio of *Aphelinus varipes* (Fürster, 1841) (Hymenoptera: Aphelinidae) varied from 92% female when reared at 25°C, to 70% at 20°C (Rohne, 2002). By contrast, in the parasitoid *Diaeretiella rapae* (M'Intosh, 1855) (Hymenoptera: Aphidiidae), the percentage of females was lowest at 26.7°C compared to 21°C (Bernal & Gonzalez, 1997). The effect of temperature on sex allocation is complicated by the fact that the sex ratio can be influenced by host size, with males more likely to emerge from smaller hosts. Host size is also

linked to temperature. For example, the body size of the black bean aphid, *Aphis fabae* Scopoli 1763, is larger when reared at low temperatures (Li & Mills, 2004).

3.2.1.4 Diapause

Diapause is an environmentally pre-programmed and hormonally mediated state of low metabolic activity associated with arrested development and increased resistance to environment extremes (Tauber, Tauber, & Masaki, 1986). Warmer and shorter winters act on the diapause strategy of insects. For example, the parasitoid *Aphidius avenae* Haliday 1834 was detected for the first time in its non-diapausing form on cereal fields in Brittany in the winter of 2011–12, whereas during the previous 20 years, it was observed in the diapausing phase outside cereal crops during winters and it only colonized the crops in spring (van Baaren et al., 2004; Krespi, Dedryver, & Creach, 1997; Le Lann et al., 2011). These modifications in the host-parasitoid relationship were linked to an increase in winter temperatures (Andrade, Krespi, Bonnardot, van Baaren, & Outreman, 2016). This shift in diapause strategy modified the food web, with *A. avenae* becoming a dominant species in this system and with an increase in the parasitism rate in winter. The plasticity of diapause induction may allow an efficient exploitation of available resources and, consequently, non-diapausing parasitoids should only be associated with anholocyclic aphids (i.e. aphids with incomplete life cycle, or which do not alternate parthenogenesis and sexual reproduction) that remain reproductive throughout winter. More generally, the parasitoid should remain synchronized with the host cycle (Godfray, 1994). The parasitoid either follows the diapause strategy of its host, and in this case the parasitoid does not kill its host before entering diapause, or the diapause strategy is decoupled from the host. In the context of climate change, the challenge for the parasitoid is to remain synchronized with the host strategy and to avoid the end of its diapause when the host is still diapausing.

3.2.1.5 Endosymbionts

Both parasitoids and herbivore insects may host endosymbiotic bacteria that can influence various life-history traits (for a review see chapter: Influence of Microbial Symbionts on Plant–Insect Interactions by Giron et al., 2016). For example, aphids have a variety of bacterial symbionts including an obligate association with the mutualist *Buchnera aphidicola* (Shigenobu, Watanabe, Hattori, Sakaki, & Ishikawa, 2000). They may also possess

secondary symbionts (e.g., *Wolbachia*) which are conditionally beneficial or deleterious (Degnanj & Moran, 2008). These bacteria have major effects on aphid biology, including enhanced tolerance to heat stress and changes in host plant range (Moran, Degnan, Santos, Dunbar, & Ochman, 2005; Tsuchida, Koga, & Fukatsu, 2004). Both major groups of endosymbionts, *Wolbachia* and *Buchnera*, may be negatively affected or eliminated by short exposures to high temperature (Thomas & Blanford, 2003). Moreover, the effect of secondary endosymbionts varies with temperature. Endosymbionts protect the aphid *Acyrtosiphum pisum* against heat stress at 25°C, whereas they decrease the fecundity at 20°C (Cheng, Montllor, & Purcell, 2000). The defensive immunity conferred to aphids by the endosymbiont *Candidatus Hamiltonella defensa* fails under heat stress (Bensadja, Boudreault, Guaya, Michaud, & Cloutier, 2006). In conclusion, climate change is expected to have broad-ranging indirect impacts on insect communities via direct effects on their associated endosymbionts.

3.2.1.6 Virus Transmission

Climate change is expected to modify the incidence of phytophagous-borne virus infections since temperature influences virus transmission (Thomas & Blanford, 2003). For example, barley yellow dwarf (BYD) is one of the world's most severe viral diseases of autumn-sown cereals. BYD viruses are transmitted by several aphid species. Environmental conditions govern the pattern and timing of primary infections by viruliferous alates and the speed of subsequent virus dissemination by apterae (secondary spread). Temperature influences both the population dynamics of the vector (e.g., 5°C is the developmental threshold for the aphid *Rhopalosiphum padi* (Linnaeus, 1758)) and the virus transmission process (e.g., 15°C is the take-off threshold for alatae). Warm January to August periods generate a high percentage of viruliferous aphids the following autumn, perhaps due to an increased rate of population growth and virus transmission in agroecosystems (Fabre et al., 2005).

3.2.2 Synchronization Between Trophic Levels

3.2.2.1 Phenological Synchronization

Numerous studies have reported climate change-induced phenological modifications at one particular trophic level. But few studies quantified the consequences of phenological shifts at several trophic levels. In UK, *Anthocharis cardamines* (Linneaus, 1758) (Lepidoptera: Pieridae) exactly follows the phenology of its host plant, for which the phenology is

advanced by 2–3 weeks with warming, but this example is an exception (Parmesan, 2006). The phenological synchrony between plants and insects, and between hosts and parasitoids, can be decoupled if the species are affected by temperature differently. For example, Harrington et al. (2001) showed in the system *Picea sitchensis* Carrière (Pinaceae)–*Operophtera brumata* (Linnaeus, 1758) (Lepidoptera: Geometridae) that the onset of bud development is not modified by warming, whereas the emergence date of the insects that feed on them is advanced. Nevertheless, the development of the caterpillars collapses because buds are not yet available when they emerge.

Few studies focused on the second and third trophic levels, and they suggest that their synchrony could be affected too. Annual variation in phenological asynchrony may be an important destabilizing factor (Godfray, Hassell, & Holt, 1994). Similarly, the time of parasitoid arrival in an aphid population influences the growth rate of the parasitoid population and its impact on the host population (Hoover & Newman, 2004). If the parasitoid arrives too early (before the aphid population has entered the exponential growth phase), the parasitoid population disappears before the aphid population increases or the parasitoid eliminates the small aphid population before it reaches the exponential growth phase. By contrast, if the parasitoid arrives during the exponential growth phase of the host, large parasitoid populations will be produced. Even small climatic changes can affect the synchrony of parasitoid activity with host populations, with large effects on the population dynamics of both.

3.2.2.2 Spatial Synchronization

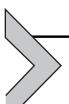
The geographical distribution of plants, herbivores and parasitoids may change during climate change as a result of various processes, including changes in the dispersal ability of insects and their host plants, and the destabilization of resident ecosystems (Parmesan, 2006). In the last century, the geographical distribution of species was modified by climate changes in 75% and 81% of the studied species in tropical and temperate areas, respectively (Parmesan, 1996). Phytophagous species often extend their geographic distribution northwards in temperate regions following an increase in temperature (Parmesan, 2006). For example, *Pararge aegeria* (Linnaeus, 1758) (Lepidoptera: Nymphalidae) extended its repartition towards the North since 1940 (Hill, Thomas, & Huntley, 1999). Extending the distribution becomes easier when the phytophagous insect succeeds to adapt to a new host plant. Also, a phytophagous species extending its

repartition can reach zones which are free of any natural enemies, as it is the case for the processionary pine moth, *Thaumetopoea pityocampa* (Denis & Schiffermüller, 1775) (Battisti et al., 2005).

3.2.3 Impacts at the Guild or at the Community Levels

Communities are mixtures of specialist and generalist species. It is predicted that specialists will suffer from climate change more than generalists, because generalists exploit a large variety of host species which can be adapted to different climatic conditions (Stireman et al., 2005). The trophic rank hypothesis predicts that higher-trophic level organisms are more negatively affected by environmental changes and disturbance than species in the lower trophic levels (Holt, Lawton, Polis, & Martinez, 1999; Tscharntke & Brandl, 2004). Moreover, at each trophic level, species succeed each other along the season, or they can be active at the same time and compete with each other. Climate change can affect the phenology of species and then the strength of the competition between species.

For phytophagous insects exploiting different plants along the season, or for natural enemies exploiting different host/prey along the season, the success requires the temporal synchronization between the different species. Changes in temperature can modify the degree to which this synchronization occurs. For example, the ant *Iridomyrmex humilis* (Mayr, 1868) (Hymenoptera: Dolichoderinae) is parasitized by several parasitoids which appear successively during the season. The first species is active below 14°C, the followers are active between 14°C and 23°C and the latest species emerges above 23°C. Temperature changes will induce discontinuity or reinforce competition between these species (Folgareit, Bruzzone, & Gilbert, 2003). However, studies at this scale are rare, impairing general conclusions.



4. IMPACT OF HUMAN ACTIVITIES ON PLANT–INSECT INTERACTIONS

4.1 The Main Anthropogenic Drivers Affecting Plant–Insect Interactions

Here, we focus on the ecological and evolutionary effects of anthropogenic factors on plant–insect interactions. We suggest a distinction between two categories of human influences: acts on purpose and acts by accident.

4.1.1 Effects of Human Eco-Engineering

4.1.1.1 Influence of Agrosystems

Agriculture is mankind's great project from the Neolithic Revolution. Creating agro-ecosystems means reducing ecosystems to what is required for the primary production of biomass: an edible plant in a favourable environment. Agrosystems differ from ecosystems in their instability and the high density of plants in monospecific stands. In addition, agrosystems show rapid and contrasting changes of the milieu, at least alternating bare soil and synchronized plant growths. Agrosystems are also characterized by the weakness of physical and chemical defences of plants towards the herbivores (Chen, Gols, & Benrey, 2015).

Agriculture and forestry select pest insects that are adapted to these agricultural conditions, in particular to the abundance and short-time availability of food resources (Bianchi, Booij, & Tscharntke, 2006; Carnus et al., 2006). Insect pests usually combine high fertility and multivoltinism with great voracity and ability to access the crops. These adaptations contribute to a high population growth rate. Half of the million described species of insects are herbivorous. Among them, around 10,000 species are pests (Herrera & Pellmyr, 2002). Pests are rather rare in natural ecosystems (Pimentel et al., 1992) because herbivorous insect populations are generally regulated by the quantity and accessibility of the plants that they exploit. But 'natural pests' exist too. For example, they are represented by crickets (Orthoptera: Acrididae) in dry tropical ecosystems. Pest insects may devastate temperate and Nordic forests, for example the oak processionary, *T. processionea*. Existence of these 'natural pests' can be explained by the similarity between these ecosystems and some agrosystems: shortness of plant growth periods in the dry tropics, homogeneous forest stands over large surfaces in cold regions (Schvester, 1985).

Agriculture causes biogeographical disruptions. Thus, it is a source of sympatric or ecological speciation (Orr & Smith, 1998). Crop plants are established anywhere possible until they become cosmopolitan (wheat, rice, corn etc.). Therefore plant crops come into contact with new insects, and new trophic relationships can take place. Adaptations of insects to new plant hosts are considered as the key mechanism of sympatric speciation. The first case of speciation via host plant shift was described by Walsch around 1860 (Berlocher & Feder, 2002). The fruit fly *Rhagoletis pomonella* Walsh 1867 (Diptera: Tephritidae) shifted from the hawthorn, *Crataegus* sp. (Rosaceae), to the common apple tree, *Malus pumila* (Rosaceae). Agriculture and the transport of plants and insects amplify this type of

diversification. This is probably why examples of sympatric diversification are less observed in natural ecosystems. For example, the European species *Ostrinia nubilalis* (Hübner, 1796) (Lepidoptera: Crambidae) recently split into two sympatric sibling species which eat different wild plants ([Malusa, Delacky, et al., 2007](#); [Malusa, Leniaud, et al., 2007](#)).

4.1.1.2 Making Landscapes From Ecosystems

Humans destroy natural habitats to build new anthropized environments, the landscapes. The main ecosystem that was destroyed in Europe was forest ([Klemm, 1996](#), pp. 17–34). In ecology, landscapes are often analyzed as systems deviating from the original ecosystem. They are characterized by their deviation ‘distance’, which integrates the difference between landscape areas (open spaces) and relics of forests (woodlots), or a proportion of wooded areas and the degree of their connections (wooded hedgerows) compared to open spaces. This metric has led to major developments in modelling ([With & Crist, 1995](#)), in particular by making the analogy between the fragmented ecosystem and island biogeography ([Fahrig, 2003](#); [Ricketts, 2001](#); [Young, Boyle, & Brown, 1996](#)), and by the duality in the landscape between cultivated and uncultivated areas ([Carré et al., 2009](#)). However, authors such as [Turner \(2005\)](#) criticized the reference to the original ecosystem, represented by the less anthropic environments, which prevents considering the landscape itself as a new ecosystem ([Manel, Schwartz, Luikart, & Taberlet, 2003](#)). Landscapes are more heterogeneous than the original systems. They abound in contact zones between anthropic and semi-natural environments. This is very conducive to the diversification of relations between insects and plants.

4.1.1.3 Urbanization Effects

The urban environment can be considered as an extreme form of landscape, often with no precise boundary between the city and rural landscapes ([McKinney, 2002](#)). Even the most densely urbanized areas offer opportunities for spontaneous life which represent, however, small biomass. Small and interstitial habitats (borders, wasteland), and mini-landscapes (parks, cemeteries) and particular niches may host adapted or synanthropic species (commensal or parasitic). Cities are places where cultivated exotic plants abound, although native plants are surprisingly well represented ([Frankie & Ehler, 1978](#); [Kühn, Brandl, & Klotz, 2004](#)) despite the difficulty to access soil, water and light. Plant–insect relationships are subjected to the same evolutionary influences than in other anthropized environments, but the communities and ecological

conditions differ. The herbivorous insects that are tolerated by their host plants in natural landscapes become multivoltine pests in urban areas, such as the aphid *Aphis nerii* Boyer de Fonscolombe 1841 on the shrub *Nerium oleander* L. (Apocynaceae) (Frankie & Ehler, 1978).

4.1.2 Evolutionary Responses to Accidental Anthropogenic Drivers

4.1.2.1 The Effects of Non-Intentional Anthropogenic Drivers

Human actions have secondary, not intended outcomes with effects without apparent spatial relationship with their cause. Unintentional disturbances are mainly changes in environmental conditions (pollution, dust, night lighting, urbanization etc.) and community (introductions and depopulation). This phenomenon is best illustrated by the historical example known under the generic term ‘industrial melanism of the peppered moth, *Biston betularia*’. In the 1950s, the British biologist Kettlewell worked on the assumption of an ecological factor of selection specific to industrialized regions and which should favour the black form of the butterfly rather than the white form. He discovered that the mortality of the white form was greatly increased because they were more visible to birds on trees darkened by industrial dust (Grant, Owen, & Clarke, 1996; Howlett & Majerus, 1987).

4.1.2.2 Community Disturbances (Introductions, Population Declines and Species Extinctions)

Species introductions are a worldwide problem. Human activities multiply the transportation of species (Kenis, Rabitsch, Auger-Rozenberg, & Roques, 2007). Symmetrically, we observe populations declining or disappearing, and extinction of species. In Europe, insect introductions accelerated from 4 introductions per year between 1971 and 1990 to 11 between 1991 and 2007, while higher plant introductions decreased from 27 to 18 during the same period (Hulme, Pysek, Nentwig, & Vila, 2009). Part of these taxa settle down on arrival, either through their pre-adaptations or because they are human commensals (cultivated plants and their pests, honeybees etc.). Introduced species have to contest a specific niche to indigenous species, but they take advantage of losing their main competitors in the native ecosystem (Bossdorf et al., 2005). The introduction, however, corresponds to a genetic bottleneck (Estoup et al., 2016). Some herbivore insect populations have a genetic trait that makes them invasive, and sometimes this trait is linked to a single gene (Lee, 2002). Thus, the invasive aphid *Acyrtosiphum pisum* loose the sexual part of its life cycle, thereby gaining the ability to switch to new host plants (Via, 2001).

Species extinction is a major concern in biodiversity conservation. Declines and extinction can trigger concatenated disturbances because of the interdependence of species in ecosystems. It is certainly the case for species associated in the mutualism of the entomophilous pollination (Berenbaum et al., 2006). For example, the decline of wild bees (non-*Apis*) is linked to the decline of some entomogamous flowering plants (Biesmeijer et al., 2006). In Europe, an unknown factor, likely of anthropogenic origin, is widely unfavourable to the biodiversity of pollinators and flowering plants.

4.2 Impact of Pollution on Plant–Insect Interactions

The number of pollutants is high and the plant–insect responses are highly diverse. Here, we address the major common characteristics that govern the bottom-up (host plant quality) and the top-down (natural enemies) processes that influence the fitness and population dynamics of herbivore insects. Inorganic and organic pollutants include air pollutants such as ozone, sulphur oxides (SO_x), nitrogen oxides (NO_x), carbon oxides (CO_x), fluorides and acidic precipitations, as well as soil pollutants such as metalloids and heavy metals (Butler & Trumble, 2008). Pollutants are more or less toxic to the environment according to their chemical composition, and this toxicity depends on the dose (concentration \times time) (see Calatayud et al., 2013 for review). Indeed, pollutants can have more impact on the environment when applied for a short period than when they are present for a long time, at equal dose. In plants, this peak effect is explained by plant response time relative to the pollutant residence time: plants do not have time to initiate their defence systems towards pollutants.

After penetration of the pollutant into plant tissues via the stomata or the roots, the pollutant generates a stress, such as oxidative stress for oxide pollutants and disturbance of calcium metabolism in the case of fluorides pollution (Calatayud, Garrec, & Nicole, 2013). Classically, the plant limits the absorption of pollutant and increases tolerance to it by implementing physical processes (e.g., stomatal closure, falling leaves) as well as chemical and biochemical processes (e.g., production of insoluble precipitates, enzymatic degradation by P450). The plant resistance to pollutants depends on the combination of the defence mechanisms already present in plant tissues and the defence processes that the plant can activate after the stress occurs. Resistance, however, also depends on other abiotic factors (e.g., temperature, humidity, light) and biotic factors (e.g., age, disease, genotype), which can have positive or negative impacts on plant response to air and soil pollution. *In situ*, there is generally an increase in insect populations

on plants in polluted areas through both bottom-up and top-down processes (Hain, 1987).

4.2.1 Bottom–Up Effects of Pollution

Pollutants have a crucial impact on the key parameters of plants that govern plant–insect relationships by changing the physiology and the biochemistry of plant tissues (Nicole, 2002). In particular, pollutants alter the parameters related to recognition by insects, nutritional quality and plant defences.

4.2.1.1 Location and Recognition of Plants

Pollutants can cause changes in the colour of plants, thereby influencing the colour of associated insects. The most famous example was reported above by the melanism mutation in British peppered moths during the industrial revolution and recently shown to be due to a transposable element (van't Hof et al., 2016). In addition, pollutants disrupt the chemical communication in plant–insect relationships. In general, the alteration of VOCs emission occurs in polluted plants, impacting orientation behaviour in herbivore insects (Blande, Holopainen, & Niinemets, 2014). Pollutants act on the plant physiology (e.g., limiting volatile emissions by stomatal closure, modifying the plant physico-chemical characteristics), or by direct degradation of VOCs in the air.

4.2.1.2 Nutritional Quality of Plants

Accumulations of toxic pollutants in plant organs such as heavy metals, arsenic and fluorine are often the cause of poisoning of plant-eating insects (Führer, 1985). In addition, pollution leads to changes in primary and secondary metabolites in plants. In particular, there is often an increase in leaf concentration of amino acids (proline), soluble protein and sugars, thereby increasing the nutritional quality of plants for insects (Kainulainen, Holopainen, & Holopainen, 2000; Warrington, 1989). Indeed, pollutants such as SO₂ and NO_x increase the concentrations of S and N in plants, with positive effects on insects. This is especially true for plants on roadsides which are major sources of NO_x (Bolsinger & Flückiger, 1987; Braun & Flückiger, 1985). But conversely, CO₂ pollution associated with NO_x leads to lower nitrogen concentrations in leaves with negative consequences for herbivore insects (Bezemer & Jones, 1998; Hättenschwiler & Schaffner, 1999). Heavy metals have in general a negative impact on the fitness of herbivores (Butler & Trumble, 2008).

4.2.1.3 Changes in Chemical and Physical Plant Defences

If secondary compounds (e.g., phenolic compounds) are part of the chemical defence systems of plants, the cuticle in turn becomes an effective barrier to insects. Pollutants such as ozone and CO₂ induce oxidative stress, and are the source of increased concentrations of phenolic compounds in plants with a negative impact on the nutritional quality of leaves for insects (Bolsinger, Lier, & Hughes, 1992; Bolsinger, Lier, Lansky, & Hughes, 1991). Meanwhile, ozone and elevated CO₂ promote the production of cuticular waxes, improving the characteristics of the physical barrier of the cuticle (Percy et al., 2002). In addition, the fitness of herbivores declines when they feed on plants contaminated by metalloids such as selenium and fluoride (Butler & Trumble, 2008).

4.2.2 Top–Down and Guild Effects of Pollution

Little is known about the impacts of pollutants on natural enemies, compared to herbivore insects. Butler and Trumble (2008) highlighted likely trends of pollution effects on insect parasitoids, predators and pathogens. The most common responses were either no effects or negative effects on natural enemies. This was reported in environments with elevated CO₂, ozone, heavy metals, metalloids and acidic precipitation. Butler and Trumble (2008) found that foliage feeders and miners often exhibit negative responses to pollutants through bottom-up process in CO₂ and heavy metal enriched environments. By contrast, phloem feeders exhibit greater fitness in environments polluted by SO₂ and NO₂. No conclusive pattern of pollution effects on bottom-up or top-down processes can be done on xylem feeders, mesophyll feeders, seed feeders and gallers; and no information is available on the fitness of borers, root feeders and, more surprisingly, pollinators (Butler & Trumble, 2008).



5. CONCLUSION AND PERSPECTIVES

The complex interactions between direct and indirect effects of global change makes particularly difficult to predict its net impact on plant–insect relationships (Fig. 1). The complexity of the mechanisms at play combines with the extreme level of diversification of life-history traits in both plants and insects. For example, insects demonstrate a high diversity of feeding modes which transposes into a high variability of plant eco-physiological responses to herbivory (Welter, 1989). Another layer of variability should be expected on the effects of global change on these specific plant responses,

as reflected by the difficulty to generalize on the impacts of temperature and CO₂ on insect herbivores (Zvereva & Kozlov, 2006). Therefore, it seems illusive to extract a single general scheme depicting the net effect of global change on plant–insect relationships.

The intricate network of direct and indirect impacts of global change on plant–insect relationships is probably even more complex than it seems because several influential processes are still understudied. For example, the biochemical pathways of heat tolerance in plants are relatively well understood (Wahid, Gelani, Ashraf, & Foolad, 2007), but it remains to elucidate if these paths interconnect with the chemical machinery involved in plant defences against herbivore insects. A link can be expected because several molecules and chemicals are involved in the two processes, such as abscisic and salicylic acids, ethylene or phenolic compounds like flavonoids. In addition, pollutants may modulate these links between plant heat tolerance and plant defences. Pollutants decrease the ability of insects to tolerate heat (Slotsbo et al., 2009), but it is not clear if the same effect applies to plants.

Urban systems may provide good models to study these complex relationships because trophic webs are simplified, and also because cities may simulate the future abiotic conditions for natural habitats (Youngsteadt, Dale, Terando, Dunn, & Frank, 2014). Urban areas are often warmer and drier on average compared to natural ecosystems (Oke, 1982). The spatial heterogeneity in surface temperature in urban zones can be used as a ‘laboratory’ to study climate change impacts on the behaviour of insects (Pincebourde et al., 2016). Given the numerous stressors experienced by plants in cities, it seems appropriate to explore the complex links between plant tolerance to heat and drought, plant defences to herbivores and emission of VOC by plants in cities. However, we are not aware of such studies. Recently, it was observed that scale insects were more abundant on trees in cities as a direct effect of the urban heat island (Youngsteadt et al., 2014), suggesting that tree species may be less protected from pests in urban areas.

Nowadays, humans develop methods to increase the resilience of plant–insect communities to global changes, in particular in the agronomic context. Among them, the Conservation Biological Control (CBC, see Chapter 10 for a review) increases the efficiency of natural enemies and decreases the impacts of pests through modification of the environment (Eilenberg, Hajek, & Lomer, 2001). The success of the CBC, however, requires good knowledge of the plant–insect interactions. Several studies have shown that more diverse plant communities are functionally less susceptible

to environmental stress (Steudel et al., 2012). This emphasizes the need to maintain biodiversity as an insurance against impacts of changing environmental conditions and sets the stage for exploring the mechanisms underlying biodiversity effects in stressed ecosystems. Indeed, biodiversity has unique effects such as complementarities and redundancies among species, dilution of plant species, or improving the stability of food webs (McCann, 2000; Naeem, 1998). Higher plant diversity, through association of plant species supplying alternative food and/or shelters at different seasons of the year, can increase the performance and fitness of natural enemies (Bompard, Jaworski, Bearez, & Desneux, 2013; Wratten, Gillespie, Decourtye, Mader, & Desneux, 2012) thus increasing their resistance to climate stresses. More diverse vegetation can promote associational resistance of host plants. This associational resistance may result from dilution of (plant) hosts for insect herbivores and from a complementarity of (insect) hosts for phytophagous-enemies, as well as a redundancy among these phytophagous-enemies. This hypothesis will be particularly important for plants whose defences are weakened by climate stress. Alternatively, the associational resistance may emerge from a decrease in climate resistance of herbivores (e.g., delayed phenology of their host plants) and an increase in climate resistance of phytophagous-enemies (e.g., climate shelters, nectar). This hypothesis will be particularly important for phytophagous insects and phytophagous-enemies under climate stress.

ACKNOWLEDGEMENTS

We warmly thank the volume editors Nicolas Sauvion, Paul-André Calatayud and Denis Thiéry for giving us the opportunity to contribute. This work was partially supported by the grant ANR Blanc MicroCliMite (ANR-2010 BLAN-1706-02) to SP, and by a Swiss National Science Foundation grant 31003A_159869 to SR.

REFERENCES

- Andrade, T. O., Krespi, L., Bonnardot, V., van Baaren, J., & Outreman, Y. (2016). Impact of climate conditions on the diversity and function of a guild of aphid parasitoids. *Oecologia*, 180, 877–888.
- Angilletta, M. J. (2009). *Thermal adaptation—A theoretical and empirical synthesis*. Oxford, UK: Oxford University Press.
- van Baaren, J., Héterier, V., Hance, T., Krespi, L., Cortesero, A.-M., Poinsot, D., ... Outreman, Y. (2004). Playing the hare or the tortoise in parasitoids: Could different oviposition strategies have an influence in host partitioning in two *Aphidius* species? *Ethology, Ecology and Evolution*, 16, 231–242.
- Bale, J. S., Masters, G. J., Hodgkinson, I. D., Awmack, C., Bezemer, T. M., Brown, V. K., ... Whittaker, J. B. (2002). Herbivory in global climate change research: Direct effects of rising temperature on insect herbivores. *Global Change Biology*, 8, 1–16.

- Barbehenn, R. V., Karowe, D. N., & Chen, Z. (2004). Performance of a generalist grasshopper on a C₃ and a C₄ grass: Compensation for the effects of elevated CO₂ on plant nutritional quality. *Oecologia*, 140, 96–103.
- Battisti, A., Stastny, M., Netherer, S., Robinet, C., Schopf, A., Roques, A., ... Larsson, S. (2005). Expansion of geographic range in the pine processionary moth caused by increased winter temperatures. *Ecological Applications*, 15, 2084–2096.
- Bensadia, F., Boudreault, S., Guaya, F., Michaud, D., & Cloutier, C. (2006). Aphid clonal resistance to a parasitoid fails under heat stress. *Journal of Insect Physiology*, 52, 146–157.
- Berenbaum, M., Bernhardt, P., Buchmann, S., Calderone, N. W., Goldstein, P., Inouye, D. W., ... Thompson, F. C. (2006). *Status of pollinators in North America*. Washington, DC, USA: National Academic Press.
- Berlocher, S. H., & Feder, J. L. (2002). Sympatric speciation in phytophagous insects: Moving beyond controversy? *Annual Review of Entomology*, 47, 773–815.
- Bernal, J., & Gonzalez, D. (1997). Reproduction of *Diaeretiella rapae* on Russian wheat aphid hosts at different temperatures. *Entomologia Experimentalis et Applicata*, 82, 159–166.
- Bezemer, T. M., & Jones, T. H. (1998). Plant–insect herbivore interactions in elevated atmospheric CO₂: Quantitative analyses and guild effects. *Oikos*, 82, 212–222.
- Bianchi, F. J. J. A., Booij, C. J. H., & Tscharntke, T. (2006). Sustainable pest regulation in agricultural landscapes: A review on landscape composition, biodiversity and natural pest control. *Proceedings of the Royal Society of London B*, 273, 1715–1727.
- Bidart-Bouzat, M. G., & Imeh-Nathaniel, A. (2008). Global change effects on plant chemical defenses against insect herbivores. *Journal of Integrative Plant Biology*, 50, 1339–1354.
- Bidart-Bouzat, M. G., Mithen, R., & Berenbaum, M. R. (2005). Elevated CO₂ influences herbivory-induced defense responses of *Arabidopsis thaliana*. *Oecologia*, 145, 415–424.
- Biesmeijer, J. C., Roberts, S. P. M., Reemer, M., Ohlemüller, R., Edwards, M., Peeters, T., ... Settele, J. (2006). Parallel declines in pollinators and insect-pollinated plants in Britain and The Netherlands. *Science*, 313, 351–354.
- Björkman, C., & Niemelä, P. (2015). *Climate change and insect pests*. London, UK: CABI Climate Change Series, CAB International.
- Blande, J. D., Holopainen, J. K., & Ninemets, U. (2014). Plant volatiles in a polluted atmosphere: Stress response and signal degradation. *Plant, Cell & Environment*, 37, 1892–1904.
- Bolsinger, M., & Flückiger, W. (1987). Enhanced aphid infestation at motorways: The role of ambient air pollution. *Entomologia Experimentalis et Applicata*, 45, 237–243.
- Bolsinger, M., Lier, M. E., & Hughes, P. R. (1992). Influence of ozone air pollution on plant–herbivore interactions. Part 2: Effects of ozone on feeding preference, growth and consumption rates of monarch butterflies (*Danaus plexippus*). *Environmental Pollution*, 77, 31–37.
- Bolsinger, M., Lier, M. E., Lansky, D. M., & Hughes, P. R. (1991). Influence of ozone air pollution on plant–herbivore interactions. Part 1: Biochemical changes in ornamental milkweed (*Asclepias curassavica* L.; asclepiadaceae) induced by ozone. *Environmental Pollution*, 72, 69–83.
- Bompard, A., Jaworski, C. C., Bearez, P., & Desneux, N. (2013). Sharing a predator: Can an invasive alien pest affect the predation on a local pest? *Population Ecology*, 55, 433–440.
- Bossdorf, O., Auge, H., Lafuma, L., Rogers, W. E., Siemann, E., & Prati, D. (2005). Phenotypic and genetic differentiation between native and introduced plant populations. *Oecologia*, 144, 1–11.
- Braun, S., & Flückiger, W. (1985). Increased population of the aphid *Aphis pomi* at a motorway. Part 3 – The effect of exhaust gases. *Environmental Pollution*, 56, 209–216.
- Browse, J., & Howe, G. A. (2008). New weapons and a rapid response against insect attack. *Plant Physiology*, 146, 832–838.

- Burrows, M. T., Schoeman, D. S., Buckley, L. B., Moore, P., Poloczanska, E. S., Brander, K. M., ... Holding, J. (2011). The pace of shifting climate in marine and terrestrial ecosystems. *Science*, 334, 652–655.
- Buse, A., & Good, J. E. G. (1996). Synchronisation of larval emergence in winter moth (*Operophtera brumata* L.) and budburst in pedunculate oak (*Quercus robur* L.) under simulated climate change. *Ecological Entomology*, 21, 335–343.
- Butler, C. D., & Trumble, J. T. (2008). Effects of pollutants on bottom-up and top-down processes in insect-plant interactions. *Environmental Pollution*, 156, 1–10.
- Caillou, R., Suppo, C., Casas, J., Woods, H. A., & Pincebourde, S. (2014). Warming decreases thermal heterogeneity of leaf surfaces: Implications for behavioural thermoregulation by arthropods. *Functional Ecology*, 28, 1449–1458.
- Calatayud, P.-A., Garrec, J.-P., & Nicole, M. (2013). Adaptation des plantes aux stress environnementaux. In N. Sauvion, P.-A. Calatayud, D. Thiéry, & F. Marion-Poll (Vol. Eds.), *Interactions insectes—plantes* (pp. 229–245). Paris: Co-edition IRD-Quae.
- Carnus, J. M., Parrotta, J., Brockerhoff, E., Arbez, M., Jactel, H., Kremer, A., ... Walters, B. (2006). Planted forests and biodiversity. *Journal of Forestry*, 104, 65–77.
- Carré, G., Roche, P., Chifflet, R., Morison, N., Bommarco, R., Harrison-Cripps, J., ... Settele, J. (2009). Landscape context and habitat type as drivers of bee biodiversity in European annual crops. *Agriculture, Ecosystems and Environment*, 133, 40–47.
- Chen, Y. H., Gols, R., & Benrey, B. (2015). Crop domestication and its impact on naturally selected trophic interactions. *Annual Review of Entomology*, 60, 35–58.
- Cheng, D. Q., Montllor, C. B., & Purcell, A. H. (2000). Fitness effects of two facultative endosymbiotic bacteria on the pea aphid, *Acyrthosiphon pisum*, and the blue alfalfa aphid, *A. kondoi*. *Entomologia Experimentalis et Applicata*, 95, 315–323.
- Coley, P. (1998). Possible effects of climate change on plant/herbivore interactions in moist tropical forests. *Climatic Change*, 39, 455–472.
- Coley, P. D., Bryant, J. P., & Chapin, F. S. (1985). Resource availability and plant antiherbivore defense. *Science*, 230, 895–899.
- Cook, G. D., Dixon, J. R., & Leopold, A. C. (1964). Transpiration: Its effects on plant leaf temperature. *Science*, 144, 546–547.
- Cornelissen, T. (2011). Climate change and its effects on terrestrial insects and herbivory patterns. *Neotropical Entomology*, 40, 155–163.
- Cornelissen, T., Stiling, P., & Drake, B. (2003). Elevated CO₂ decreases leaf fluctuating asymmetry and herbivory by leaf miners on two oak species. *Global Change Biology*, 10, 27–36.
- Coviella, C. E., & Trumble, J. T. (1999). Effects of elevated atmospheric carbon dioxide on insect-plant interactions. *Conservation Biology*, 13, 700–712.
- Curran, E. D., Labandeira, C. C., & Wilf, P. (2010). Fossil insect folivory tracks paleotemperature for six million years. *Ecological Monographs*, 80, 547–567.
- Degnanj, P. H., & Moran, N. A. (2008). Evolutionary genetics of a defensive facultative symbiont of insects: Exchange of toxin-encoding bacteriophage. *Molecular Ecology*, 17, 916–929.
- Dicke, M., & Baldwin, I. T. (2010). The evolutionary context for herbivore-induced plant volatiles: Beyond the ‘cry for help’. *Trends in Plant Science*, 15, 167–175.
- Doane, J. F., Lee, Y. W., Klingler, J., & Westcott, N. D. (1975). Orientation response of *Ctenicera destructor* and other wireworms (Coleoptera: Elateridae) to germinating grain and to carbon dioxide. *Canadian Entomologist*, 107, 1233–1252.
- Docherty, M., Wade, F. A., Hurst, D. K., Whittaker, J. B., & Lea, P. J. (1997). Responses of tree sap-feeding herbivores to elevated CO₂. *Global Change Biology*, 3, 51–59.

- Duchateau, M. J., Hoshiba, H., & Velthuis, H. H. W. (1994). Diploid males in the bumble bee *Bombus terrestris*, sex determination, sex alleles and viability. *Entomologia Experimentalis et Applicata*, 71, 263–269.
- Dury, S. J., Good, J. E. G., Perrins, C. M., Buse, A., & Kaye, T. (1998). The effects of increasing CO₂ and temperature on oak leaf palatability and the implications for herbivorous insects. *Global Change Biology*, 4, 55–61.
- Dussourd, D. E., & Denno, R. F. (1991). Deactivation of plant defense: Correspondence between insect behavior and secretory canal architecture. *Ecology*, 72, 1383–1396.
- Ehleringer, J. R., Cerling, T. E., & Dearing, M. D. (2002). Atmospheric CO₂ as a global change driver influencing plant-animal interactions. *Integrative and Comparative Biology*, 42, 424–430.
- Eilenberg, J., Hajek, A., & Lomer, C. (2001). Suggestions for unifying the terminology in biological control. *BioControl*, 46, 387–400.
- Estoup, A., Ravigné, V., Hufbauer, R., Vitalis, R., Gautier, M., & Facon, B. (2016). Is there a genetic paradox of biological invasion? *Annual Review of Ecology, Evolution, and Systematics*, 47. <http://dx.doi.org/10.1146/annurev-ecolsys-121415-032116>.
- Fabre, F., Plantegenest, M., Mieuget, L., Dedryver, C. A., Leterrier, J. L., & Jacquot, E. (2005). Effects of climate and land use on the occurrence of viruliferous aphids and the epidemiology of barley yellow dwarf disease. *Agriculture, Ecosystems and Environment*, 106, 49–55.
- Facey, S. L., Ellsworth, D. S., Staley, J. T., Wright, D. J., & Johnson, S. N. (2014). Upsetting the order: How climate and atmospheric change affects herbivore–enemy interactions. *Current Opinion in Insect Science*, 5, 66–74.
- Fahrig, L. (2003). Effects of habitat fragmentation on biodiversity. *Annual Review of Ecology, Evolution, and Systematics*, 34, 487–515.
- Fajer, E. D., Bowers, M. D., & Bazzaz, F. A. (1991). The effects of enriched CO₂ atmospheres on the buckeye butterfly, *Junonia coenia*. *Ecology*, 72, 751–754.
- Farmer, E. E., Almíreas, E., & Krishnamurthy, V. (2003). Jasmonates and related oxylipins in plant responses to pathogenesis and herbivory. *Current Opinion in Plant Biology*, 6, 372–378.
- Feyereisen, R. (1999). Insect P450 enzymes. *Annual Review of Entomology*, 44, 507–533.
- Fine, P. V. A., Mesones, I., & Coley, P. D. (2004). Herbivores promote habitat specialization by trees in amazonian forests. *Science*, 305, 663–665.
- Folgareit, P. J., Bruzzone, O. A., & Gilbert, L. E. (2003). Seasonal patterns of activity among species of black fire ant parasitoid flies (*Pseudacteon*: Phoridae) in Argentina explained by analysis of climatic variables. *Biological Control*, 28, 368–378.
- Frankie, G. W., & Ehler, L. E. (1978). Ecology of insects in urban environments. *Annual Review of Entomology*, 23, 367–387.
- Franzén, M., & Ockinger, E. (2012). Climate-driven changes in pollinator assemblages during the last 60 years in an Arctic mountain region in Northern Scandinavia. *Journal of Insect Conservation*, 16, 227–238.
- Führer, E. (1985). Air pollution and the incidence of forest insect problems. *Zeitschrift für Angewandte Entomologie*, 99, 371–377.
- Futuyma, D. J., & Agrawal, A. A. (2009). Macroevolution and the biological diversity of plants and herbivores. *Proceedings of the National Academy of Sciences of the United States of America*, 106, 18054–18061.
- Gates, D. M. (1980). *Biophysical ecology*. New York, USA: Springer-Verlag.
- Ghasemzadeh, A., Jaafar, H. Z., & Rahmat, A. (2010). Elevated carbon dioxide increases contents of flavonoids and phenolic compounds, and antioxidant activities in Malaysian young ginger (*Zingiber officinale* Roscoe) varieties. *Molecules*, 15, 7907–7922.
- Giron, D., Dedeine, F., Dubreuil, G., Huguet, E., Mouton, L., Outreman, Y., Vavre, F., & Simon, J.-C. (2016). Influence of microbial symbionts on plant–insect interactions. In

- N. Sauvion, D. Thiery, & P. A. Calatayud (Eds.), *Insect-Plant Interactions* (Vol. 81). (in this volume).
- Godfray, H. C. J. (1994). *Parasitoids: Behavioural and evolutionary ecology*. Princeton, New Jersey, USA: Princeton University Press.
- Godfray, H. C. J., Hassell, M. P., & Holt, R. D. (1994). The population dynamic consequences of phenological asynchrony between parasitoids and their hosts. *Journal of Animal Ecology*, 63, 1–10.
- Graham, R. W., & Grimm, E. C. (1990). Effects of global climate change on the patterns of terrestrial biological communities. *Trends in Ecology & Evolution*, 5, 289–292.
- Grant, B. S., Owen, D. F., & Clarke, C. A. (1996). Parallel rise and fall of melanic peppered moths in America and Britain. *Journal of Heredity*, 87, 351–357.
- Guenther, A. B., Zimmerman, P. R., Harley, P. C., Monson, R. K., & Fall, R. (1993). Isoprene and monoterpene emission rate variability: Model evaluations and sensitivity analyses. *Journal of Geophysical Research*, 98, 12609–12617.
- Hain, F. P. (1987). Interactions of insects, trees and air pollutants. *Tree Physiology*, 3, 93–102.
- Hance, T., van Baaren, J., Vernon, P., & Boivin, G. (2007). Impact of extreme temperatures on parasitoids in a climate change perspective. *Annual Review of Entomology*, 52, 107–126.
- Harrington, R., Fleming, R. A., & Woiwod, I. P. (2001). Climate change impacts on insect management and conservation in temperate regions: Can they be predicted? *Agricultural and Forest Meteorology*, 3, 233–240.
- Harrington, R., Woiwod, I., & Sparks, T. (1999). Climate change and trophic interactions. *Trends in Ecology & Evolution*, 14, 146–150.
- Hättenschwiler, S., & Schhäuser, C. (1999). Opposing effects of elevated CO₂ and N deposition on *Lymantria monacha* larvae feeding on spruce trees. *Oecologia*, 118, 210–217.
- Heckel, D.G. (2014). Insect detoxification and sequestration strategies. In C. Voelckel, & G. Jander (Vol. Eds.), *Annual plant reviews: Vol. 47. Insect-plant interactions* (pp. 77–114). Chichester, UK: John Wiley & Sons, Ltd. <http://dx.doi.org/10.1002/9781118829783.ch3>.
- Heinrich, B. (1979). *Bumblebee economics*. Cambridge, London, England: Harvard University Press.
- Heinrich, B., & Buchmann, S. L. (1986). Thermoregulatory physiology of the carpenter bee, *Xylocopa varipuncta*. *Journal of Comparative Physiology B*, 156, 557–562.
- Helliker, B. R., & Richter, S. L. (2008). Subtropical to boreal convergence of tree-leaf temperatures. *Nature*, 454, 511–515.
- Herms, D. A., & Mattson, W. J. (1992). The dilemma of plants – to grow or defend. *Quarterly Review of Biology*, 67, 283–335.
- Herrera, C. M., & Pellmyr, O. (2002). *Plant – Animal interactions. An evolutionary approach*. Oxford, UK: Blackwell Science.
- Hewitt, G. (1999). Post-glacial re-colonization of European biota. *Biological Journal of the Linnean Society*, 68, 87–112.
- Hill, J. K., Thomas, C. D., & Huntley, B. (1999). Climate and habitat availability determine 20th century changes in a butterfly's range margin. *Proceedings of the Royal Society of London B*, 266, 1197–1206.
- van't Hof, A. E., Campagne, P., Rigden, D. J., Yung, C. J., Lingley, J., Quail, M. A., ... Saccheri, I. J. (2016). The industrial melanism mutation in British peppered moths is a transposable element. *Nature*, 534, 102–105.
- Holt, R. D., Lawton, J. H., Polis, G. A., & Martinez, N. D. (1999). Trophic rank and the species-area relationship. *Ecology*, 80, 1495–1504.
- Hoover, J. K., & Newman, J. A. (2004). Tritrophic interactions in the context of climate change: A model of grasses, cereal aphids and their parasitoids. *Global Change Biology*, 10, 1197–1208.

- Howlett, R. J., & Majerus, M. E. N. (1987). The understanding of industrial melanism in the peppered moth (*Biston betularia*) (Lepidoptera: Geometridae). *Biological Journal of the Linnean Society*, 30, 31–44. <http://dx.doi.org/10.1111/j.1095-8312.1987.tb00286.x>.
- Huey, R. B., & Stevenson, R. D. (1979). Integrating physiology and ecology of ectotherms: A discussion of approaches. *American Zoologist*, 19, 357–366.
- Hulme, P. E., Pysek, P., Nentwig, W., & Vila, M. (2009). Will threat of biological invasions unite the European Union? *Science*, 324, 40–41.
- Hunter, M. D. (2001). Effects of elevated atmospheric carbon dioxide on insect–plant interactions. *Agricultural and Forest Entomology*, 3, 153–159.
- IPCC (2007). Contribution of Working Group I to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change. In S. Solomon, D. Qin, M. Manning, M. Marquis, K. Averyt, M.M.B. Tignor, ..., Z. Chen (Eds.), *Climate Change 2007: The Physical Science Basis* Cambridge, UK.
- IPCC. (2014). *Climate change 2014: Impacts, adaptation, and vulnerability*. Cambridge, UK: Cambridge University Press.
- Jaenike, J. (1990). Host specialization in phytophagous insects. *Annual Review of Ecology, Evolution, and Systematics*, 21, 243–273.
- Jamieson, M. A., Trowbridge, A. M., Raffa, K. F., & Lindroth, R. L. (2012). Consequences of climate warming and altered precipitation patterns for plant–insect and multitrophic interactions. *Plant Physiology*, 160, 1719–1727.
- Jepsen, J. U., Hagen, S. B., Ims, R. A., & Yoccoz, N. G. (2008). Climate change and outbreaks of the geometrids *Operophtera brumata* and *Epiprilla autumnata* in subarctic birch forest: Evidence of a recent outbreak range expansion. *Journal of Animal Ecology*, 77, 257–264.
- Johns, C. V., Beaumont, L. J., & Hughes, L. (2003). Effects of elevated CO₂ and temperature on development and consumption rates of *Octotoma championi* and *O. scabripennis* feeding on *Lantana camara*. *Entomologia Experimentalis et Applicata*, 108, 169–178.
- Johns, C. V., & Hughes, L. (2002). Interactive effects of elevated CO₂ and temperature on the leaf-miner *Dialectica scalariella* Zeller (Lepidoptera: Gracillariidae) in Paterson's Curse, *Echium plantagineum* (Boraginaceae). *Global Change Biology*, 8, 142–152.
- Julkunen-Tiitto, R., Nybakken, L., Randriamanana, T., & Virjamo, V. (2015). Boreal woody species resistance affected by climate change. In C. Björkman, & P. Niemelä (Vol. Eds.), *Climate change and insect pests* (pp. 54–73). Wallingford: CABI.
- Kainulainen, P., Holopainen, J. K., & Holopainen, T. (2000). Combined effect of ozone and nitrogen on secondary compounds, amino acids, and aphid performance in Scots pine. *Journal of Environmental Quality*, 29, 334–342.
- Kaiser, L., Ode, P., van Nouhuys, S., Calatayud, P.-A., Colazza, S., Cortesero, A.-M., Thiel, A., & van Baaren, J. (2016). The Plant as a habitat for entomophagous insects. In N. Sauvion, D. Thiery, & P. A. Calatayud (Eds.), *Insect-Plant Interactions* (Vol. 81). (in this volume).
- Kaspari, M., Clay, N. A., Lucas, J., Yanoviak, S. P., & Kay, A. (2015). Thermal adaptation generates a diversity of thermal limits in a rainforest ant community. *Global Change Biology*, 21, 1092–1102.
- Kenis, M., Rabitsch, W., Auger-Rozenberg, M.-A., & Roques, A. (2007). How can alien species inventories and interception data help us prevent in sect invasions? *Bulletin of Entomological Research*, 97, 489–502.
- Kergoat, G. J., Meseguer, A. S., & Jousselin, E. (2016). Evolution of plant–insect interactions: insights from macroevolutionary approaches in plants and herbivorous insects. In N. Sauvion, D. Thiery, & P. A. Calatayud (Eds.), *Insect-Plant Interactions* (Vol. 81). (in this volume).
- Kerr, E. D., Phelan, C., & Woods, H. A. (2013). Subtle direct effects of rising atmospheric CO₂ on insect eggs. *Physiological Entomology*, 38, 302–305.

- Kerr, J. T., Pindar, A., Galpern, P., Packer, L., Potts, S. G., Roberts, S. M., ... Wagner, D. L. (2015). Climate change impacts on bumblebees converge across continents. *Science*, *349*, 177–180.
- Kessler, A., & Heil, M. (2011). The multiple faces of indirect defences and their agents of natural selection. *Functional Ecology*, *25*, 348–357.
- Klemm, M. (1996). Man-made bee habitats in the anthropogenous landscape of central Europe – Substitutes for threatened or destroyed riverine habitats?. In *Linnean society symposium series* (Vol. 18) Academic Press Limited.
- Körner, C. (2007). The use of altitude in ecological research. *Trends in Ecology & Evolution*, *22*, 569–574.
- Krespi, L., Dedryver, C.-A., & Creach, V. (1997). Variability in the development of cereal aphid parasitoids and hyperparasitoids in oceanic regions as a response to climate and abundance of hosts. *Environmental Entomology*, *26*, 545–551.
- Kühn, L., Brandl, R., & Klotz, S. (2004). The flora of German cities is naturally species rich. *Evolutionary Ecology Research*, *6*, 749–764.
- Le Lann, C., Roux, O., Serain, N., van Alphen, J. J. M., Vernon, P., & van Baaren, J. (2011). Thermal tolerance of sympatric hymenopteran parasitoid species: Does it match seasonal activity? *Physiological Entomology*, *36*, 21–28.
- Lecocq, T., Dellicour, S., Michez, D., Lhomme, P., Vanderplanck, M., Valterova, I., ... Rasmont, P. (2013). Scent of a break-up: Phylogeography and reproductive trait divergences in the red-tailed bumblebee (*Bombus lapidarius*). *BMC Evolutionary Biology*, *13*, 263.
- Lee, C. E. (2002). Evolutionary genetics of invasive species. *Trends in Ecology & Evolution*, *17*, 386–391.
- Li, B. P., & Mills, N. (2004). The influence of temperature on size as an indicator of host quality for the development of a solitary koinobiont parasitoid. *Entomologia Experimentalis et Applicata*, *110*, 249–256.
- Lieutier, F., Bermudez-Torres, K., Cook, J., Harris, M. O., Legal, L., Sallé, A., Schatz, B., & Giron, D. (2016). From plant exploitation to mutualism. In N. Sauvion, D. Thiery, & P. A. Calatayud (Eds.), *Insect-Plant Interactions* (Vol. 81). (in this volume).
- Linacre, E. T. (1967). Further notes on a feature of leaf and air temperatures. *Theoretical and Applied Climatology*, *15*, 422–436.
- Lincoln, D. E., Fajer, E. D., & Johnson, R. H. (1993). Plant-insect herbivore interactions in elevated CO₂ environments. *Trends in Ecology & Evolution*, *8*, 64–68.
- Lindroth, R. L. (2010). Impacts of elevated atmospheric CO₂ and O₃ on forests: Phytochemistry, trophic interactions, and ecosystem dynamics. *Journal of Chemical Ecology*, *36*, 2–21.
- Loreto, F., & Schnitzler, J.-P. (2010). Abiotic stresses and induced BVOCs. *Trends in Plant Science*, *15*, 154–166.
- Malusa, T., Dalecky, A., Ponsard, S., Audiot, P., Streiff, R., Chaval, Y., & Bourguet, D. (2007). Genetic structure and gene flow in French populations of two *Ostrinia* taxa: Host races or sibling species? *Molecular Ecology*, *16*, 4210–4222.
- Malusa, T., Leniaud, L., Martin, J.-F., Audiot, P., Bourguet, D., Ponsard, S., ... Dopman, E. (2007). Molecular differentiation at nuclear loci in French host races of the European corn borer (*Ostrinia nubilalis*). *Genetics*, *176*, 2343–2355.
- Manel, S., Schwartz, M. K., Luikart, G., & Taberlet, P. (2003). Landscape genetics: Combining landscape ecology and population genetics. *Trends in Ecology & Evolution*, *18*, 189–197.
- Marchal, P., & Feytaud, J. (1911). On the parasite of the eggs of the grape berry moth and the vine moth. *Comptes Rendus Hebdomadaires des Séances de l'Académie des Sciences*, *153*, 633–636.

- Martin-Vertedor, D., Ferrero-Garcia, J. J., & Torres-Vila, L. M. (2010). Global warming affects phenology and voltinism of *Lobesia botrana* in Spain. *Agricultural and Forest Entomology*, 12, 169–176.
- Martinet, B., Lecocq, T., Smet, J., & Rasmont, P. (2015). A protocol to assess insect resistance to heat waves, applied to bumblebees (*Bombus Latreille*, 1802). *PLoS One*, 10, e0118591.
- Martinet, B., Rasmont, P., Cederberg, B., Evrard, D., Ødegård, F., Paukkunen, J., & Lecocq, T. (2016). Forward to the North: Two Euro-Mediterranean bumblebee species now cross the Arctic Circle. *Annales de la Société entomologique de France (N.S.)*, 51, 303–309.
- Mason, P. G., & Hopper, K. R. (1997). Temperature dependence in locomotion of the parasitoid *Aphelinus asychis* (Hymenoptera: Aphelinidae) from geographical regions with different climates. *Environmental Entomology*, 26, 1416–1423.
- May, M. L. (1979). Insect thermoregulation. *Annual Review of Entomology*, 24, 313–349.
- McCann, K. S. (2000). The diversity–stability debate. *Nature*, 405, 228–233.
- McKinney, M. L. (2002). Urbanization, biodiversity, and conservation. *BioScience*, 52, 883–890.
- Meehl, G. A., & Tebaldi, C. (2004). More intense, more frequent and longer lasting heat waves in the 21st century. *Science*, 305, 994–997.
- Michaletz, S. T., Weiser, M. D., Zhou, J., Kaspari, M., Helliker, B. R., & Enquist, B. J. (2015). Plant thermoregulation: Energetics, trait–environment interactions, and carbon economics. *Trends in Ecology & Evolution*, 30, 714–724.
- Michener, C. D. (1979). Biogeography of the bees. *Annals of the Missouri Botanical Garden*, 66, 277–342.
- Michener, C. D. (2007). *The bees of the world* (2nd ed.). Baltimore, USA: The Johns Hopkins University.
- Mitter, C., Farrell, B., & Futuyma, D. J. (1991). Phylogenetic studies of insect-plant interactions: Insights into the genesis of diversity. *Trends in Ecology & Evolution*, 6, 290–293.
- Moran, N. A., Degnan, P. H., Santos, S. R., Dunbar, H. E., & Ochman, H. (2005). The players in a mutualistic symbiosis: Insects, bacteria, viruses, and virulence genes. *Proceedings of the National Academy of Sciences of the United States of America*, 102, 16919–16926.
- Morison, J. I. L., & Lawlor, D. W. (1999). Interactions between increasing CO₂ concentration and temperature on plant growth. *Plant, Cell & Environment*, 22, 659–682.
- Nabity, P. D., Hillstrom, M. L., Lindroth, R. L., & DeLucia, E. H. (2012). Elevated CO₂ interacts with herbivory to alter chlorophyll fluorescence and leaf temperature in *Betula papyrifera* and *Populus tremuloides*. *Oecologia*, 169, 905–913.
- Naeem, S. (1998). Species redundancy and ecosystem reliability. *Conservation Biology*, 12, 39–45.
- Nicolas, G., & Sillans, D. (1989). Immediate and latent effects of carbon dioxide on insects. *Annual Review of Entomology*, 34, 97–116.
- Nicole, M. C. (2002). Les relations des insectes phytophages avec leurs plantes hôtes. *Antennae*, 9, 6.
- Nieto, A., Roberts, S. P. M., Kemp, J., Rasmont, P., Kuhlmann, M., García Criado, M., ... De Meulemeester, T. (2014). *European red list of bees*. Brussels: International Union for Conservation of Nature, 03/2015; Publication Office of the European Union.
- Ode, P. J., Johnson, S. N., & Moore, B. D. (2014). Atmospheric change and induced plant secondary metabolites — Are we reshaping the building blocks of multi-trophic interactions? *Current Opinion in Insect Science*, 5, 57–65.
- Oke, T. R. (1982). The energetic basis of the urban heat island. *Quarterly Journal of the Royal Meteorological Society*, 108, 1–24.
- Orr, M. R., & Smith, T. B. (1998). Ecology and speciation. *Trends in Ecology & Evolution*, 16, 502–506.

- Owen, E. L., Bale, J. S., & Hayward, S. A. L. (2013). Can winter-active bumblebees survive the cold? Assessing the cold tolerance of *Bombus terrestris* audax and the effects of pollen feeding. *PLoS One*, 8(11), e80061.
- Oyen, J. O., Susma, G., & Dillon, M. E. (2016). Altitudinal variation in bumblebee (*Bombus*) critical thermal limits. *Journal of Thermal Biology*, 59, 52–57.
- Parmesan, C. (1996). Climate and species' range. *Nature*, 382, 765–766.
- Parmesan, C. (2006). Ecological and evolutionary responses to recent climate change. *Annual Review of Ecology, Evolution, and Systematics*, 37, 637–669.
- Parmesan, C., & Yohe, G. (2003). A globally coherent fingerprint of climate change impacts across natural systems. *Nature*, 421, 37–42.
- Pateman, R. M., Hill, J. K., Roy, D. B., Fox, R., & Thomas, C. D. (2012). Temperature-dependent alterations in host use drive rapid range expansion in a butterfly. *Science*, 336, 1028–1030.
- Patiny, S., & Michez, D. (2007). Biogeography of bees (Hymenoptera, Apoidea) in Sahara and the Arabian deserts. *Insect Systematics & Evolution*, 38, 19–34.
- Pauli, H., Gottfried, M., Dullinger, S., Abdaladze, O., Akhalkatsi, M., Alonso, J. L. B., ... Ghosn, D. (2012). Recent plant diversity changes on Europe's mountain summits. *Science*, 336, 353–355.
- Pellissier, L., Fiedler, K., Ndribe, C., Dubuis, A., Pradervand, J.-N., Guisan, A., & Rasmann, S. (2012). Shifts in species richness, herbivore specialization, and plant resistance along elevation gradients. *Ecology and Evolution*, 2, 1818–1825.
- Pellissier, L., Moreira, X., Danner, H., Serrano, M., Salamin, N., van Dam, N. M., & Rasmann, S. (2016). The simultaneous inducibility of phytochemicals related to plant direct and indirect defences against herbivores is stronger at low elevation. *Journal of Ecology*, 104, 1116–1125.
- Pellissier, L., Roger, A., Bilat, J., & Rasmann, S. (2014). High elevation *Plantago lanceolata* plants are less resistant to herbivory than their low elevation conspecifics: Is it just temperature? *Ecography*, 37, 950–959.
- Peñuelas, J., & Llusia, J. (2003). BVOCs: Plant defense against climate warming? *Trends in Plant Science*, 8, 105–109.
- Peñuelas, J., & Staudt, M. (2010). BVOCs and global change. *Trends in Plant Science*, 15, 133–144.
- Percy, K. E., Awmack, C. S., Lindroth, R. L., Kubiske, M. E., Kopper, B. J., Isebrands, J. G., ... Oksanen, E. (2002). Altered performance of forest pests under atmospheres enriched by CO₂ and O₃. *Nature*, 420, 403–407.
- Pimentel, D., Stachow, U., Takacs, D. A., Brubaker, H. W., Dumas, A. R., Meaney, J. J., ... Corzilius, D. B. (1992). Conserving biological diversity in agricultural/forestry systems. *BioScience*, 42, 354–362.
- Pincebourde, S., & Casas, J. (2006a). Leaf miner-induced changes in leaf transmittance cause variations in insect respiration rates. *Journal of Insect Physiology*, 52, 194–201.
- Pincebourde, S., & Casas, J. (2006b). Multitrophic biophysical budgets: Thermal ecology of an intimate herbivore insect–plant interaction. *Ecological Monographs*, 76, 175–194.
- Pincebourde, S., & Casas, J. (2015). Warming tolerance across insect ontogeny: Influence of joint shifts in microclimates and thermal limits. *Ecology*, 96, 986–997.
- Pincebourde, S., & Casas, J. (2016). Hypoxia and hypercarbia in endophagous insects: Larval position in the plant gas exchange network is key. *Journal of Insect Physiology*, 84, 137–153.
- Pincebourde, S., Frak, E., Sinoquet, H., Régnard, J. L., & Casas, J. (2006). Herbivory mitigation through increased water use efficiency in a leaf mining moth–apple tree relationship. *Plant, Cell & Environment*, 29, 2238–2247.

- Pincebourde, S., Murdock, C. C., Vickers, M., & Sears, M. W. (2016). Fine-scale microclimatic variation can shape the responses of organisms to global change in both natural and urban environments. *Integrative and Comparative Biology*, icw016.
- Pincebourde, S., Sinoquet, H., Combes, D., & Casas, J. (2007). Regional climate modulates the canopy mosaic of favourable and risky microclimates for insects. *Journal of Animal Ecology*, 76, 424–438.
- Pincebourde, S., & Suppo, C. (2016). The vulnerability of tropical ectotherms to warming is modulated by the microclimatic heterogeneity. *Integrative and Comparative Biology*, 56, 85–97.
- Pincebourde, S., & Woods, H. A. (2012). Climate uncertainty on leaf surfaces: The biophysics of leaf microclimates and their consequences for leaf-dwelling organisms. *Functional Ecology*, 26, 844–853.
- Polce, C., Garratt, M. P., Termansen, M., Ramirez-Villegas, J., Challinor, A. J., Lappage, M. G., ... Somerwill, K. E. (2014). Climate-driven spatial mismatches between British orchards and their pollinators: Increased risks of pollination deficits. *Global Change Biology*, 20, 2815–2828.
- Potter, K. A., Davidowitz, G., & Woods, H. A. (2009). Insect eggs protected from high temperatures by limited homeothermy of plant leaves. *Journal of Experimental Biology*, 212, 3448–3454.
- Proctor, M., Yeo, P., & Lack, A. (1996). *The natural history of pollination*. Portland, Oregon, USA: Timber Press Inc.
- Rasmann, S., Alvarez, N., & Pellissier, L. (2014a). The altitudinal niche-breadth hypothesis in insect-plant interactions. *Annual Plant Reviews*, 47, 338–359.
- Rasmann, S., & Pellissier, L. (2015). Adaptive responses of plants to insect herbivores under climate change. In C. Björkman, & P. Niemelä (Vol. Eds.), *Climate change and insect pests* (pp. 38–53). Wallingford, UK: CABI.
- Rasmann, S., Pellissier, L., Defossez, E., Jactel, H., & Kunstler, G. (2014b). Climate-driven change in plant–insect interactions along elevation gradients. *Functional Ecology*, 28, 46–54.
- Rasmont, P., Franzen, M., Lecocq, T., Harpke, A., Roberts, S. P. M., Biesmeijer, K., ... Schweiger, O. (2015). Climatic risk and distribution atlas of European bumblebees. *Biorisk*, 10, 1–246. <http://dx.doi.org/10.3897/biorisk.10.4749>.
- Rasmont, P., & Iserbyt, S. (2012). The bumblebees scarcity syndrome: Are heat waves leading to local extinctions of bumblebees (Hymenoptera: Apidae: *Bombus*)? *Annales de la Société Entomologique de France (N.S.)*, 48, 275–280.
- Reddy, G. V. P., Holopainen, J. K., & Guerrero, A. (2002). Olfactory responses of *Plutella xylostella* natural enemies to host pheromone, larval frass, and green leaf cabbage volatiles. *Journal of Chemical Ecology*, 28, 131–143.
- Ricketts, T. H. (2001). The matrix matters: Effective isolation in fragmented landscapes. *American Naturalist*, 158, 87–99.
- Robinson, E. A., Ryan, G. D., & Newman, J. A. (2012). A meta-analytical review of the effects of elevated CO₂ on plant–arthropod interactions highlights the importance of interacting environmental and biological variables. *New Phytologist*, 194, 321–336.
- Robinson, P. J. (2001). On the definition of a heat wave. *Journal of Applied Meteorology*, 40, 762–775.
- Rohne, O. (2002). Effect of temperature and host stage on performance of *Aphelinus varipes* Forster (Hym., Aphelinidae) parasitizing the cotton aphid, *Aphis gossypii* Glover (Hom., Aphididae). *Journal of Applied Entomology*, 126, 572–576.
- Romero, G. Q., & Koricheva, J. (2011). Contrasting cascade effects of carnivores on plant fitness: A meta-analysis. *Journal of Animal Ecology*, 80, 696–704.

- Rosenthal, G. A., & Berenbaum, M. R. (1991). *Herbivores: Their interactions with secondary plant metabolites*. San Diego, USA: Academic Press, Inc.
- Rundle, H. D., & Nosal, P. (2005). Ecological speciation. *Ecology Letters*, 8, 336–352.
- Salt, D. T., Brooks, G. L., & Whittaker, J. B. (1995). Elevated carbon dioxide affects leaf-miner performance and plant growth in docks (*Rumex spp.*). *Global Change Biology*, 1, 153–156.
- de Sassi, C., Staniczenko, P. P. A., & Tylianakis, J. M. (2012). Warming and nitrogen affect size structuring and density dependence in a host-parasitoid food web. *Philosophical Transactions of the Royal Society B*, 367, 3033–3041.
- Saudreau, M., Pincebourde, S., Dassot, M., Adam, B., Loxdale, H. D., & Biron, D. G. (2013). On the canopy structure manipulation to buffer climate change effects on insect herbivore development. *Trees, Structure and Function*, 27, 239–248.
- Schmitz, O. J., Hamback, P. A., & Beckerman, A. P. (2000). Trophic cascades in terrestrial systems: A review of the effects of carnivore removals on plants. *American Naturalist*, 155, 141–153.
- Schoonhoven, L. M., van Loon, J. J. A., & Dicke, M. (2005). *Insect-plant biology*. Oxford, UK: Oxford University Press.
- Schvester, D. (1985). Regard sur la santé de nos forêts. *Revue Forestière Française* (Sp. Regard sur la santé de nos forêts), 45–64.
- Settele, J., Hammel, V., Hulme, P., Karlson, U., Klotz, S., Kotarac, M., ... Peterson, K. (2005). ALARM: Assessing large-scale environmental risks for biodiversity with tested methods. *Gaia-Ecological Perspectives for Science and Society*, 14, 69–72.
- Shigenobu, S., Watanabe, H., Hattori, M., Sakaki, Y., & Ishikawa, H. (2000). Genome sequence of the endocellular bacterial symbiont of aphids *Buchnera* sp. APS. *Nature*, 407, 81–86.
- Slotsbo, S., Heckmann, L. H., Damgaard, C., Roelofs, D., de Boer, T., & Holmstrup, M. (2009). Exposure to mercury reduces heat tolerance and heat hardening ability of the springtail *Folsomia candida*. *Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology*, 150, 118–123.
- Smith, P. H. D., & Jones, T. H. (1998). Effects of elevated CO₂ on the chrysanthemum leaf-miner, *Chromatomyia syngenesiae*: A greenhouse study. *Global Change Biology*, 4, 287–291.
- Stange, E. E., & Ayres, M. P. (2001). *Climate change impacts: insects*. eLS, John Wiley & Sons, Ltd.
- Steudel, B., Hector, A., Friedl, T., Löfke, C., Lorenz, M., Wesche, M., & Kessler, M. (2012). Biodiversity effects on ecosystem functioning change along environmental stress gradients. *Ecology Letters*, 15, 1397–1405.
- Stiling, P., & Cornelissen, T. (2007). How does elevated carbon dioxide (CO₂) affect plant–herbivore interactions? A field experiment and meta-analysis of CO₂-mediated changes on plant chemistry and herbivore performance. *Global Change Biology*, 13, 1823–1842.
- Stireman, J. O., Dyer, L. A., Janzen, D. H., Singer, M. S., Lill, J. T., Marquis, R. J., ... Barone, J. A. (2005). Climatic unpredictability and parasitism of caterpillars: Implications of global warming. *Proceedings of the National Academy of Sciences of the United States of America*, 102, 17384–17387.
- Strong, D. R., Lawton, J. H., & Southwood, R. (1984). *Insects on plants: Community patterns and mechanisms*. London, UK: Blackwell Scientific.
- Sun, Y., Yin, J., Cao, H., Li, C., Kang, L., & Ge, F. (2011). Elevated CO₂ influences nematode-induced defense responses of tomato genotypes differing in the JA pathway. *PLoS One*, 6, e19751.
- Sunday, J. M., Bates, A. E., Kearney, M. R., Colwell, R. K., Dulvy, N. K., Longino, J. T., & Huey, R. B. (2014). Thermal safety margins and the necessity of thermoregulatory

- behavior across latitude and elevation. *Proceedings of the National Academy of Sciences of the United States of America*, 111, 5610–5615.
- Tauber, M. J., Tauber, C. A., & Masaki, S. (1986). *Seasonal adaptations of insects*. New York, USA: Oxford University Press.
- Terzo, M., & Rasmont, P. (2014). *Atlas of the European bees: genus Xylocopa* (STEP Project, Atlas Hymenoptera, Mons, Gembloux).
- Thaler, J. S., Agrawal, A. A., & Halitschke, R. (2010). Salicylate-mediated interactions between pathogens and herbivores. *Ecology*, 91, 1075–1082.
- Thaler, J. S., Humphrey, P. T., & Whiteman, N. K. (2012). Evolution of jasmonate and salicylate signal crosstalk. *Trends in Plant Science*, 17, 260–270.
- Thies, C., Haenke, S., Scherber, C., Bengtsson, J., Bommarco, R., Clement, L. W., ... Hawro, V. (2011). The relationship between agricultural intensification and biological control: Experimental tests across Europe. *Ecological Applications*, 21, 2187–2196.
- Thomas, M. B., & Blanford, S. (2003). Thermal biology in insect-parasite interactions. *Trends in Ecology & Evolution*, 18, 344–350.
- Tougeron, K., van Baaren, J., Burel, F., & Alford, L. (2016). Comparing thermal tolerance across contrasting landscapes: First steps towards understanding how landscape management could modify ectotherm thermal tolerance. *Insect Conservation and Diversity*, 9, 171–180.
- Trotta, V., Calboli, F. C., Ziosi, M., Guerra, D., Pezzoli, M. C., David, J. R., & Cavicchi, S. (2006). Thermal plasticity in *Drosophila Melanogaster*: A comparison of geographic populations. *BMC Evolutionary Biology*, 6, 67.
- Tscharntke, T., & Brandl, R. (2004). Plant-insect interactions in fragmented landscapes. *Annual Review of Entomology*, 49, 405–430.
- Tsuchida, T., Koga, R., & Fukatsu, T. (2004). Host-plant specialization governed by facultative symbiont. *Science*, 303, 1989.
- Turner, M. G. (2005). Landscape ecology: What is the state of the science? *Annual Review of Ecology, Evolution, and Systematics*, 36, 319–344.
- Tylianakis, J. M., Didham, R. K., Bascompte, J., & Wardle, D. A. (2008). Global change and species interactions in terrestrial ecosystems. *Ecology Letters*, 11, 1351–1363.
- Via, S. (2001). Sympatric speciation in animals: The ugly duckling grows up. *Trends in Ecology & Evolution*, 16, 381–390.
- Virilio, P. (2005). *L'accident originel*. Paris, France: Galilée.
- Wahid, A., Gelani, S., Ashraf, M., & Foolad, M. R. (2007). Heat tolerance in plants: An overview. *Environmental and Experimental Botany*, 61, 199–223.
- Warrington, S. (1989). Ozone enhances the growth rate of cereal aphids. *Agriculture, Ecosystems and Environment*, 26, 65–68.
- Welter, S. C. (1989). Arthropod impact on plant gas exchange. *Insect-Plant Interactions*, 1, 135–150.
- Whitehorn, P. R., Tinsley, M. C., Brown, M. J. F., Darvill, B., & Goulson, D. (2009). Impacts of inbreeding on bumblebee colony fitness under field conditions. *BMC Evolutionary Biology*, 9, 152.
- Whittaker, J. B. (2001). Insects and plants in a changing atmosphere. *Journal of Ecology*, 89, 507–518.
- Williams, R. S., Norby, R. J., & Lincoln, D. E. (2000). Effects of elevated CO₂ and temperature-grown red and sugar maple on gypsy moth performance. *Global Change Biology*, 6, 685–695.
- With, K. A., & Crist, T. O. (1995). Critical thresholds in species' responses to landscape structure. *Ecology*, 76, 2446–2459.
- Woods, H. A. (2013). Ontogenetic changes in the body temperature of an insect herbivore. *Functional Ecology*, 27, 1322–1331.

- Woods, H. A., Bonnecaze, R. T., & Zrubek, B. (2005). Oxygen and water flux across egg-shells of *Manduca sexta*. *Journal of Experimental Biology*, 208, 1297–1308.
- Wratten, S. D., Gillespie, M., Decourtey, A., Mader, E., & Desneux, N. (2012). Pollinator habitat enhancement: Benefits to other ecosystem services. *Agriculture, Ecosystems and Environment*, 159, 112–122.
- Wu, J., & Baldwin, I. T. (2010). New insights into plant responses to the attack from insect herbivores. *Annual Review of Genetics*, 44, 1–24.
- Young, A., Boyle, T., & Brown, T. (1996). The population genetic consequences of habitat fragmentation for plants. *Trends in Ecology & Evolution*, 11, 413–418.
- Youngsteadt, E., Dale, A. G., Terando, A. J., Dunn, R. R., & Frank, S. D. (2014). Do cities simulate climate change? A comparison of herbivore response to urban and global warming. *Global Change Biology*, 21, 97–105.
- Zavala, J. A., Casteel, C. L., DeLucia, E. H., & Berenbaum, M. R. (2008). Anthropogenic increase in carbon dioxide compromises plant defense against invasive insects. *Proceedings of the National Academy of Sciences of the United States of America*, 105, 5129–5133.
- Zavala, J. A., Nabity, P. D., & DeLucia, E. H. (2013). An emerging understanding of mechanisms governing insect herbivory under elevated CO₂. *Annual Review of Entomology*, 58, 79–97.
- Zvereva, E. L., & Kozlov, M. V. (2006). Consequences of simultaneous elevation of carbon dioxide and temperature for plant-herbivore interactions: A meta-analysis. *Global Change Biology*, 12, 27–41.